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APPLICATION

FOR

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TITLE:

NOVEL GENES AND POLYPEPTIDES FOR THE

DIAGNOSIS OF GIANT CELL ARTERITIS

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NUCLEIC ACAS NOVEL GENES AND POLYPEPTIDES FOR THE DIAGNOSIS AND TREATMENT OF GIANT CELL ARTERITIS

FIELD OF THE INVENTION

This invention generally pertains to the fields of medicine and medical diagnostics. In particular, this invention provides novel genes and polypeptides and methods for making and using them. Specifically, the compositions and methods of the invention are used to diagnose and treat Giant Cell Arteritis (GCA).

BACKGROUND OF THE INVENTION

Giant cell arteritis (GCA) is a systemic vasculitis that is a serious and potentially blinding rheumatologic disease of the elderly. Current treatment of GCA requires systemic immunosuppression with profound morbidity in the affected elderly population. GCA is widely believed to be immune-mediated; however, the etiology and pathogenesis of this systemic vasculitis remains unidentified. Furthermore, diagnosis of GCA is difficult because it relies on a constellation of nonspecific signs and symptoms and a diagnostic arterial biopsy. Significantly, blindness may be the first symptom of GCA. Thus, if a way was found to better diagnose or even screen for early onset or predisposition for GCA at an earlier stage of the disease, many cases of blindness and many lives would be saved.

Currently, corticosteroids are critical in the treatment of giant cell arteritis; they reduce the incidence of blindness and rapidly relieve symptoms. However, the amounts of steroids (e.g., prednisone) needed are significant and not without side effects; particularly as they usually must be given over an extended period of time, usually about two years. Steroid treatment is not uniformly effective and causes significant morbidity in up to 40% of patients because of hypertension, osteoporosis, infection, glucose dysregulation, fluid overload, and aseptic necrosis of the hip or shoulder. Alternative use of nonsteroidal anti-inflammatory drugs (NSAIDs) will lessen the painful symptoms, but they do not prevent the blindness or vascular problems. Accordingly, new methods of treating GCA are needed. The present invention addresses these and other needs.

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SUMMARY OF THE INVENTION

The present invention provides novel compositions and methods in the screening for, diagnosis of and treatment of GCA.

The invention provides an isolated or recombinant nucleic acid comprising a nucleic acid sequence having at least 75% sequence identity to SEQ ID NO:1 or a nucleic acid encoding a polypeptide, wherein the polypeptide has a sequence as set forth in SEQ ID NO:2; or a nucleic acid sequence having at least 75% sequence identity to SEQ ID NO:3 or a nucleic acid encoding a polypeptide, wherein the polypeptide has a sequence as set forth in SEQ ID NO:4; or a nucleic acid sequence having at least 85% sequence identity to SEQ ID NO:5 or a nucleic acid encoding a polypeptide, wherein the polypeptide has a sequence as set forth in SEQ ID NO:6; or a nucleic acid sequence having at least 75% sequence identity to SEQ ID NO:7 or a nucleic acid encoding a polypeptide, wherein the polypeptide has a sequence as set forth in SEQ ID NO:8. In various embodiments, the sequence identity to SEQ ID NO:1 is at least 80%, 85%, 90%, 95%, and 98%; the sequence identity to SEQ ID NO:3 is at least 80%, 85%, 90%, 95%, and 98%; the sequence identity to SEQ ID NO:5 is at least 90%, 95%, and 98%; and, the sequence identity to SEQ ID NO:7 is at least 80%, 85%, 90%, 95%, and 98%. The nucleic acid can also comprises a sequence as set forth in SEQ ID NO:1; SEQ ID NO:3; SEQ ID NO:5; or SEQ ID NO:7.

The invention also provides an isolated or recombinant nucleic acid comprising a nucleic acid sequence having at least 75% sequence identity to SEQ ID NO:9, SEQ ID NO:10; SEQ ID NO:11; SEQ ID NO:13; SEQ ID NO:14, SEQ ID NO:15 or SEQ ID NO:16; or has a sequence as set forth in SEQ ID NO:12. In alternative embodiments, the sequence identity to SEQ ID NO:9, SEQ ID NO:10; SEQ ID NO:11; SEQ ID NO:13; SEQ ID NO:14, SEQ ID NO:15 or SEQ ID NO:16 is at least 80%, 85%, 90%, 95%, and 98%. The nucleic acid can also have a sequence as set forth in SEQ ID NO:9, SEQ ID NO:10; SEQ ID NO:11; SEQ ID NO:13; SEQ ID NO:14, SEQ ID NO:15 or SEQ ID NO:16.

The invention provides an isolated or recombinant nucleic acid which specifically hybridizes to a nucleic acid comprising a sequence as set forth in SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:10, SEQ ID NO:11, SEQ ID NO:13, SEQ ID NO:14, SEQ ID NO:15 or SEQ ID NO:16 under stringent conditions, wherein the stringent conditions include a wash step comprising a wash in 0.2X SSC at a temperature of about 65°C for about 15 minutes. The nucleic acid can be between about 15 and about 200

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residues in length; between about 25 and about 100 residues in length; or between about 35 and about 75 residues in length.

The invention provides an expression vector comprising at least one nucleic acid operably linked to a promoter, wherein the nucleic acid comprises a nucleic acid sequence of the invention. In the expression vector, the nucleic acid can be operably linked to the promoter in the sense orientation or the antisense orientation. Also provided is a transformed cell comprising the nucleic acids and /or expression vectors of the invention.

The invention provides a polymerase chain reaction (PCR) primer pair that can amplify a nucleic acid sequence of the invention, or a subsequence thereof, under *in situ* or *in vitro* conditions.

The invention provides an isolated or recombinantly expressed polypeptide, said polypeptide encoded by nucleic acid which specifically hybridizes to a nucleic acid comprising a sequence as set forth in SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:10, SEQ ID NO:11, SEQ ID NO:13, SEQ ID NO:14, SEQ ID NO:15 or SEQ ID NO:16 under stringent conditions, wherein the stringent conditions include a wash step comprising a wash in 0.2X SSC at a temperature of about 65°C for about 15 minutes.

The invention provides a polypeptide encoded by SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:10, SEQ ID NO:11, SEQ ID NO:12, SEQ ID NO:13, SEQ ID NO:14, SEQ ID NO:15 or SEQ ID NO:16 in any reading frame on either strand (e.g., coding strand or complementary strand); such exemplary polypeptide and peptide sequences of the invention are set forth herein.

The invention provides an isolated or recombinantly expressed polypeptide having 75% sequence identity to SEQ ID NO:4, having 85% sequence to SEQ ID NO:6 or 75% sequence identity to SEQ ID NO:8. In alternative embodiments, the polypeptide has 80%, 85%, 90%, 95%, and 98% sequence identity to an amino acid sequence as set forth in SEQ ID NO:2, SEQ ID NO:4, and SEQ ID NO:8; and, 90%, 95%, and 98% sequence identity to an amino acid sequence as set forth in SEQ ID NO:6. The isolated or recombinantly expressed polypeptide of the invention can have an amino acid sequence as set forth in SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, or SEQ ID NO:8. The isolated or recombinantly expressed polypeptide can be between about 15 and about 200 residues in length;

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between about 25 and about 100 residues in length; or between about 35 and about 75 residues in length.

The invention further provides an immunogenic peptide comprising a subsequence of a polypeptide of the invention, exemplary sequences of which are provided herein. For example, the immunogenic peptide can have a sequence as set forth from about residue 1 to residue 92 of SEQ ID NO:2; about residue 1 to 124 of SEQ ID NO:4; about residue 1 to 48 of SEQ ID NO:6; or about residue 1 to 81 of SEQ ID NO:8. The invention also provides a fusion protein comprising a polypeptide, particularly an immunogenic peptide, and a heterologous sequence. The heterologous sequence can be any sequence not GCA-associated; for example, a sequence that aids in the expression, isolation/ purification of the fusion protein.

The invention provides an isolated or recombinant antibody or binding fragment thereof which specifically binds to a polypeptide or peptide or an immunogenic fragment thereof, of the invention. The antibody can be a monoclonal antibody or a polyclonal antibody or binding fragment thereof. The invention further provides a hybridoma cell line comprising (e.g., producing) a monoclonal antibody of the invention.

The invention provides kits for detecting the presence of nucleic acid sequences associated with GCA (typically from, e.g., a serum, urine, tissue or biopsy sample) comprising a nucleic acid of the invention, or combinations thereof, where a nucleic acid in the sample detectably hybridizes to a nucleic acid of the invention under *in situ* or *in vitro* conditions. Also provided are kits for detecting the presence of nucleic acid sequences associated with GCA (also typically from, e.g., a serum, urine, tissue or biopsy sample) comprising an amplification primer pair that can amplify a sample nucleic acid having a sequence as set forth in claim 1, claim 5 or claim 9 under *in situ* or *in vitro* conditions. Further provided are kits for detecting the presence of polypeptide sequences associated with GCA comprising an antibody of the invention.

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The invention also provides kits, e.g., ELISA kits, for detecting the presence of human antibodies associated with GCA in a sample comprising a polypeptide of the invention. The polypeptides or peptides in the kit can be immobilized. The kit can further comprise a non-human antibody or an antisera that specifically binds to a human antibody under an *in situ* or *in vitro* conditions. As described below, the non-human antibody in the kit can further comprise a detectable tag (e.g., an enzyme, a radionuclide, biotin, and the like, as discussed below), or the invention can comprise a second antibody capable of binding to the first non-human antibody.

The invention provides arrays (also called "DNA chips" or "microarrays") of oligonucleotide probes for the identification of GCA-associated nucleic acid in a sample. The nucleic acid in these arrays are typically immobilized on a solid support comprising, amongst other nucleic acids, a GCA-associated nucleic acid of the invention.

The invention provides methods for diagnosing or determining predisposition for GCA comprising the following steps: (a) providing an antibody that specifically binds to a polypeptide associated with GCA, wherein the antibody has the same specificity as an antibody of the invention (that binds to a GCA-associated peptide or polypeptide); or, a nucleic acid that can detectably hybridizes to a nucleic acid of the invention under *in situ* or *in vitro* conditions; (b) providing a tissue or fluid (e.g., whole blood, serum or urine) sample; (c) contacting the antibody or nucleic acid with the sample; and (d) detecting whether the antibody specifically binds to a polypeptide in the tissue or serum sample or the nucleic acid hybridizes to a nucleic acid in the tissue or serum sample; wherein the specific binding or hybridization is diagnostic for or determines a predisposition for GCA.

The invention provides methods for diagnosing or determining predisposition for GCA comprising the following steps: (a) providing a nucleic acid amplification primer pair of the invention that can amplify a GCA-associated nucleic acid under *in situ* or *in vitro* conditions; (b) providing a tissue or fluid (e.g., whole blood, serum or urine) sample; (c) contacting the primer pair with the tissue or fluid (e.g., whole blood, serum or urine) sample under amplification reaction conditions; and (d) detecting whether the primer pair has amplified a nucleic acid in the sample; wherein hybridization is diagnostic for or determines a predisposition for GCA.

The invention provides methods for diagnosing or determining predisposition for GCA comprising the following steps: (a) providing a polypeptide or peptide of the invention (a GCA-associated polypeptide); (b) providing a tissue or fluid (e.g., whole blood, serum or urine) sample; (c) contacting the sample with the polypeptide or peptide under physiologic conditions; and (d) detecting whether an antibody in the sample specifically binds to a polypeptide or peptide of step (a); wherein antibody binding is diagnostic for or determines a predisposition for GCA. In this method the detection in step (d) can be an ELISA assay or equivalent thereof, as discussed below.

The invention provides methods for isolating nucleic acid sequences associated with GCA comprising the following steps: (a) providing a first tissue sample from a tissue or fluid

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specimen not showing histologic or other signs of GCA and a second tissue sample from a tissue or fluid specimen showing histologic or other signs of GCA; (b) isolating the nucleic acid from both samples; (c) substracting nucleic acid from the first sample from the second sample to isolate nucleic acid only present in the second sample, wherein the isolated nucleic acid from the second sample is associated with GCA-affected tissue and not normal tissue. This aspect of the invention can incorporate all variations and equivalents of substractive hybridization techniques, as described below. In this method, the first and the second tissue sections can be taken from a "skip" lesion of a temporal artery of a GCA patient.

The invention provides methods for isolating lymphocytes involved in the pathogenesis of GCA comprising the following steps: (a) incubating a GCA-associated polypeptide or peptide of the invention with a plurality of adherent, irradiated antigen presenting cell cultures; (b) contacting a sample of isolated lymphocytes from a GCA patient with the polypeptide-incubated adherent antigen presenting cell cultures of step (a); (c) culturing the cells contacted in step (b) for sufficient time to allow for cytokine secretion or cell proliferation; and (d) detecting which cell culture comprises proliferating cells or cells secreting cytokines, wherein proliferation or secretion of cytokines indicates the isolated lymphocytes are involved in the pathogenesis of GCA. In this method, the lymphocytes can be B cells, stem cells or T cells. The cells can be cultured for about 2 to 5 days before cell proliferation or cytokine secretion is analyzed.

The invention provides methods for generating antibodies for the diagnosis or treatment of GCA comprising administering a GCA-associated polypeptide or peptide of the invention in amounts sufficient to generate an immune response. The immune response can be primarily humoral, cell-based, or a combination thereof. The GCA-associated polypeptides can be administered to non-human animals to generate non-human antibodies for diagnostic tests (they can also be administered to human Ab-gene-carrying mice to generate all-human antibodies, as discussed below). The GCA-associated polypeptides also can be administered to humans as vaccines for the treatment or prevention of GCA.

The invention provides methods for making the nucleic acids, polypeptides, and antibodies of the invention, as described herein. These can be isolated from nature, made in recombinant expression systems or can be entirely synthetic, as described herein. For example, nucleic acids within the scope of the invention can be identified and isolated using, e.g., nucleic acid hybridization techniques, including *in situ* hybridization, traditional techniques such as dot-

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blotting of cDNA or genomic libraries, or amplification techniques, such as PCR. Amplification primers can be designed directly from the GCA-associated nucleic acids described herein, or they can be modified, degenerate primers, as discussed below. Polypeptides can be identified and isolated using a variety of methods, including, e.g., analysis of ORFs from GCA-associated nucleic acids, binding with GCA-associated antibodies, wherein the antibodies can be, e.g., isolated from human patients or generated using the polypeptides or peptides of the invention, and the binding can be, e.g., in tissue samples or expression libraries. Antibodies can be made by inoculation of mammalian recipients using, e.g., the polypeptides or peptides of the invention, or, they can be isolated from immunized animals or expression libraries, e.g., phage (antibody binding site) expression libraries.

A further understanding of the nature and advantages of the present invention is realized by reference to the remaining portions of the specification, the figures and claims.

All publications, patents and patent applications cited herein are hereby expressly incorporated by reference for all purposes.

BRIEF DESCRIPTION OF THE DRAWINGS

Figure 1 illustrates a representative experiment using the fusion protein GCA1b-GST in serum ELISA using various dilution of human serum from a GCA+ and GCA- individual and detecting for bound human IgG.

Figure 2 illustrates a representative experiment using the fusion protein GCA17-GST in serum ELISA using various dilution of human serum from a GCA+ and GCA- individual and detecting for bound human IgG.

Figure 3 illustrates a representative experiment using four exemplary GCA-GST fusion proteins, GCA 1a-GST, GCA 1b-GST, GCA14-GST, and GCA17-GST. These fusion proteins were used to detect serum IgG in sera from 10 GCA+ and 10 GCA- individuals.

DETAILED DESCRIPTION

The present invention provides a novel strategy for the detection, diagnosis and treatment of Giant Cell Arteritis (GCA). The invention provides for compositions and methods useful for

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early diagnosis of (including assessing the predisposition for acquiring GCA) and the treatment of GCA. Use of the invention to screen elderly patients for predisposition to GCA and early diagnosis of GCA is particularly needed to prevent a common cause of blindness in our aging population. GCA is seen especially in the elderly, usually at a mean age of about 70 years old. Blindness is the most serious and irreversible feature of GCA, where it can be sudden, painless and permanent. Involvement of both eyes can occur. Significantly, blindness may be the initial clinical presentation of GCA. Accordingly, use of the methods and composition of the invention will prevent or ameliorate (in addition to treating) significant numbers of incidents of blindness in our aging population.

The invention is based on the discovery that novel sequences can be associated with GCA lesions. While the invention is not limited by any particular theory or mechanism, these unique-GCA associated sequences may be associated with a pathology initiating or causative microorganism. Accordingly, subtractive hybridization of normal (non-involved) from GCA-involved tissue led to the discovery of the novel GCA-associated sequences of the invention. Translation of exemplary sequences to recombinant polypeptides (in the form of fusion proteins for convenience of isolation and manipulation) led to the discovery that GCA patients have circulating antibodies that specifically bind to the polypeptides of the invention. Accordingly, the peptides and polypeptides of the invention are used in kits and methods for diagnosing GCA by identifying circulating anti-GCA antibodies in the serum, urine or tissue samples of patients. Because blindness may be the first presenting symptom of GCA, the diagnostic methods of the invention can be used screen for GCA on patients that, while having no symptoms of GCA, do have a relatively high probability of suffering from GCA, such as elderly patients.

DEFINITIONS

Unless defined otherwise, all technical and scientific terms used herein have the meaning commonly understood by a person skilled in the art to which this invention belongs. As used herein, the following terms have the meanings ascribed to them unless specified otherwise.

The term "amplifying" and "amplification" as used herein incorporates its common usage and refers to the use of any suitable amplification methodology for generating or detecting recombinant or naturally expressed nucleic acid, as described in detail, below. For example, the

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invention provides methods and reagents (e.g., specific degenerate oligonucleotide primer pairs) for amplifying (e.g., by polymerase chain reaction, PCR) naturally expressed (e.g., genomic or mRNA) or recombinant (e.g., cDNA) nucleic acids of the invention (e.g., GCA-associated sequences of the invention) in vivo or in vitro.

The term "antibody" includes both intact antibodies having at least two heavy (H) chains and two light (L) chains inter-connected by disulfide bonds and antigen binding fragments thereof, either isolated from natural sources, recombinantly generated or partially or entirely synthetic. Examples of antigen binding fragments include, e.g., (i) a Fab fragment, a monovalent fragment consisting of the VL, VH, CL and CH1 domains; (ii) a F(ab')2 fragment, a bivalent fragment comprising two Fab fragments linked by a disulfide bridge at the hinge region; (iii) a Fd fragment consisting of the VH and CH1 domains; (iv) a Fv fragment consisting of the VL and VH domains of a single arm of an antibody, (v) a dAb fragment (Ward et al., (1989) Nature 341:544-546), which consists of a VH domain; and (vi) an isolated complementarity determining region (CDR). Although the two domains of the Fv fragment, VL and VH, are coded for by separate genes, they can be joined, using recombinant methods, by a synthetic linker that enables them to be made as a single protein chain in which the VL and VH regions pair to form monovalent molecules (known as single chain Fv (scFv); See e.g., Bird et al. (1988) Science 242:423-426; and Huston et al. (1988) Proc. Natl. Acad. Sci. USA 85:5879-5883). Single chain antibodies are also antibodies of the invention. Fragments can be also prepared by enzymatic or chemical cleavage of intact antibodies. "Specific binding" refers to antibody binding to a predetermined antigen. Typically, the antibody binds with an association constant (Ka) of at least about 1 x 10^6 M⁻¹ or 10^7 M⁻¹, or about 10^8 M⁻¹ to 10^9 M⁻¹, or about 10^{10} M⁻¹ to 10^{11} M⁻¹ or higher, and binds to the predetermined antigen with an affinity that is at least two-fold greater than its affinity for binding to a non-specific antigen (e.g., BSA, casein) other than the predetermined antigen or a closely-related antigen. The phrases "an antibody recognizing an antigen" and "an antibody specific for an antigen" are used interchangeably herein with the term "an antibody which binds specifically to an antigen." The term "high affinity" for an IgG antibody refers to an equilibrium association constant (K_a) of at least about 10⁷M⁻¹, at least about 10⁸M⁻¹, at least about 10⁹M⁻¹, at least about 10¹⁰M⁻¹, at least about 10¹¹M⁻¹, or at least about $10^{12} \mathrm{M}^{-1}$ or greater, e.g., up to $10^{13} \mathrm{M}^{-1}$ or $10^{14} \mathrm{M}^{-1}$ or greater. However, "high affinity" binding can vary for other antibody isotypes.

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The term "array" includes any array of probes, e.g., nucleic acids stably associated with the surface of a solid support under hybridization conditions sufficient to produce a hybridization pattern, as discussed in detail, below.

The term "expression vector" refers to any recombinant expression system for the purpose of expressing a nucleic acid sequence of the invention *in vitro* or *in vivo*, constitutively or inducibly, in any cell, including prokaryotic, yeast, fungal, plant, insect or mammalian cell. The term includes linear or circular expression systems. The term includes expression systems that remain episomal or integrate into the host cell genome. The expression systems can have the ability to self-replicate or not, *i.e.*, drive only transient expression in a cell. The term includes recombinant "expression cassettes" which contain only the minimum elements needed for transcription of the recombinant nucleic acid.

The term "fusion protein" or "fusion polypeptide" includes polypeptides having sequences which are normally unrelated to each other, e.g., a polypeptide of the invention (e.g., SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:8) linked to a "heterologous sequence" (see definition below), as discussed in detail, below.

The term "giant cell arteritis" or "GCA" describes the chronic inflammatory disease characterized by the progressive inflammation of many arteries of the body (panarteritis), and is sometimes also known as "temporal arteritis," "Horton's disease," "cranial arteritis," or "granulomatous arteritis." It is typically diagnosed by clinical symptoms and by histopathologic analysis, as described below.

The term "heterologous" when used with reference to a nucleic acid or polypeptide, indicates that a sequence that comprises two or more subsequences which are not found in the same relationship to each other as normally found in nature, or is recombinantly engineered so that its level of expression, or physical relationship to other nucleic acids or other molecules in a cell, or structure, is not normally found in nature. For instance, a heterologous nucleic acid is typically recombinantly produced, having two or more sequences from unrelated genes arranged in a manner not found in nature; *e.g.*, a nucleic acid open reading frame (ORF) of the invention operatively linked to a promoter sequence inserted into an expression cassette, e.g., a vector, of the invention. As another example, a polypeptide of the invention is linked to tag, e.g., a detection- and purification-facilitating domain, as a fusion protein.

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As used herein, the term "immune response" includes any humoral (antibody) or cellular immune response or combination thereof.

As used herein, "isolated," when referring to a molecule or composition, such as, e.g., an isolated infected cell comprising a nucleic acid sequence derived from a library of the invention, means that the molecule or composition (including, e.g., a cell) is separated from at least one other compound, such as a protein, DNA, RNA, or other contaminants with which it is associated in vivo or in its naturally occurring state. Thus, a nucleic acid or polypeptide or peptide sequence is considered isolated when it has been isolated from any other component with which it is naturally associated. An isolated composition can, however, also be substantially pure. An isolated composition can be in a homogeneous state. It can be in a dry or an aqueous solution. Purity and homogeneity can be determined, e.g., using any analytical chemistry technique, as described herein.

The term "nucleic acid" or "nucleic acid sequence" refers to a deoxy-ribonucleotide or ribonucleotide oligonucleotide, including single- or double-stranded, or coding or non-coding (e.g., "antisense") forms. The term encompasses nucleic acids, i.e., oligonucleotides, containing known analogues of natural nucleotides. The term also encompasses nucleic-acid-like structures with synthetic backbones, see e.g., Oligonucleotides and Analogues, a Practical Approach, ed. F. Eckstein, Oxford Univ. Press (1991); Antisense Strategies, Annals of the N.Y. Academy of Sciences, Vol 600, Eds. Baserga et al. (NYAS 1992); Milligan (1993) J. Med. Chem. 36:1923-1937; Antisense Research and Applications (1993, CRC Press), WO 97/03211; WO 96/39154; Mata (1997) Toxicol. Appl. Pharmacol. 144:189-197; Strauss-Soukup (1997) Biochemistry 36:8692-8698; Samstag (1996) Antisense Nucleic Acid Drug Dev 6:153-156.

As used herein, the term "operably linked," refers to a functional relationship between two or more nucleic acid (e.g., DNA) segments. Typically, it refers to the functional relationship of a transcriptional regulatory sequence to a transcribed sequence. For example, a promoter (defined below) is operably linked to a coding sequence, such as a nucleic acid of the invention, if it stimulates or modulates the transcription of the coding sequence in an appropriate host cell or other expression system. Generally, promoter transcriptional regulatory sequences that are operably linked to a transcribed sequence are physically contiguous to the transcribed sequence, i.e., they are cis-acting. However, some transcriptional regulatory sequences, such as enhancers, need not be physically contiguous or located in close proximity to the coding sequences whose

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transcription they enhance. For example, in one embodiment, a promoter is operably linked to an ORF-containing nucleic acid sequence of the invention, as exemplified by, e.g., SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:5 and SEQ ID NO:7.

The terms "polypeptide," "protein," and "peptide" include compositions of the invention that also include "analogs," or "conservative variants" and "mimetics" ("peptidomimetics") with structures and activity that substantially correspond to the exemplary sequences, such as SEQ ID NO:2; SEQ ID NO:4; SEQ ID NO:6; or SEQ ID NO:8. Thus, the terms "conservative variant" or "analog" or "mimetic" also refer to a polypeptide or peptide which has a modified amino acid sequence, such that the change(s) do not substantially alter the polypeptide's (the conservative variant's) structure and/or activity (e.g., immunogenicity, ability to bind to human antibodies, etc.), as defined herein. These include conservatively modified variations of an amino acid sequence, i.e., amino acid substitutions, additions or deletions of those residues that are not critical for protein activity, or substitution of amino acids with residues having similar properties (e.g., acidic, basic, positively or negatively charged, polar or non-polar, etc.) such that the substitutions of even critical amino acids does not substantially alter structure and/or activity. Conservative substitution tables providing functionally similar amino acids are well known in the art. For example, one exemplary guideline to select conservative substitutions includes (original residue followed by exemplary substitution): ala/gly or ser; arg/ lys; asn/ gln or his; asp/glu; cys/ser; gln/asn; gly/asp; gly/ala or pro; his/asn or gln; ile/leu or val; leu/ile or val; lys/arg or gln or glu; met/leu or tyr or ile; phe/met or leu or tyr; ser/thr; thr/ser; trp/tyr; tyr/trp or phe; val/ile or leu. An alternative exemplary guideline uses the following six groups, each containing amino acids that are conservative substitutions for one another: 1) Alanine (A), Serine (S), Threonine (T); 2) Aspartic acid (D), Glutamic acid (E); 3) Asparagine (N), Glutamine (Q); 4) Arginine (R), Lysine (K); 5) Isoleucine (I), Leucine (L), Methionine (M), Valine (V); and 6) Phenylalanine (F), Tyrosine (Y), Tryptophan (W); (see also, e.g., Creighton (1984) Proteins, W.H. Freeman and Company; Schulz and Schimer (1979) Principles of Protein Structure, Springer-Verlag). One of skill in the art will appreciate that the above-identified substitutions are not the only possible conservative substitutions. For example, for some purposes, one may regard all charged amino acids as conservative substitutions for each other whether they are positive or negative. In addition, individual substitutions, deletions or additions that alter, add or delete a single amino

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acid or a small percentage of amino acids in an encoded sequence can also be considered "conservatively modified variations."

The terms "mimetic" and "peptidomimetic" refer to a synthetic chemical compound that has substantially the same structural and/or functional characteristics of the polypeptides of the invention (e.g., ability to bind, or "capture," human antibodies in an ELISA). The mimetic can be either entirely composed of synthetic, non-natural analogues of amino acids, or, is a chimeric molecule of partly natural peptide amino acids and partly non-natural analogs of amino acids. The mimetic can also incorporate any amount of natural amino acid conservative substitutions as long as such substitutions also do not substantially alter the mimetics' structure and/or activity. As with polypeptides of the invention which are conservative variants, routine experimentation will determine whether a mimetic is within the scope of the invention, i.e., that its structure and/or function is not substantially altered. Polypeptide mimetic compositions can contain any combination of non-natural structural components, which are typically from three structural groups: a) residue linkage groups other than the natural amide bond ("peptide bond") linkages; b) non-natural residues in place of naturally occurring amino acid residues; or c) residues which induce secondary structural mimicry, i.e., to induce or stabilize a secondary structure, e.g., a beta turn, gamma turn, beta sheet, alpha helix conformation, and the like. A polypeptide can be characterized as a mimetic when all or some of its residues are joined by chemical means other than natural peptide bonds. Individual peptidomimetic residues can be joined by peptide bonds, other chemical bonds or coupling means, such as, e.g., glutaraldehyde, N-hydroxysuccinimide esters, bifunctional maleimides, N,N'-dicyclohexylcarbodiimide (DCC) or N,N'diisopropylcarbodiimide (DIC). Linking groups that can be an alternative to the traditional amide bond ("peptide bond") linkages include, e.g., ketomethylene (e.g., -C(=O)-CH₂- for -C(=O)-NH-), aminomethylene (CH₂-NH), ethylene, olefin (CH=CH), ether (CH₂-O), thioether (CH₂-S), tetrazole (CN₄-), thiazole, retroamide, thioamide, or ester (see, e.g., Spatola (1983) in Chemistry and Biochemistry of Amino Acids, Peptides and Proteins, Vol. 7, pp 267-357, "Peptide Backbone Modifications," Marcell Dekker, NY). A polypeptide can also be characterized as a mimetic by containing all or some non-natural residues in place of naturally occurring amino acid residues; non-natural residues are well described in the scientific and patent literature.

The term "pharmaceutical composition" refers to a composition suitable for pharmaceutical use in a subject. The pharmaceutical compositions of this invention are formulations that comprise a pharmacologically effective amount of a composition comprising, e.g., a nucleic acid (including, e.g., ribozymes, antisense oligonucleotides, and other inhibitory nucleic acid variations), a vector, or an antibody of the invention, and a pharmaceutically acceptable carrier.

As used herein, the term "promoter" includes all sequences capable of driving transcription of a coding sequence in an expression system. Thus, promoters used in the constructs of the invention include *cis*-acting transcriptional control elements and regulatory sequences that are involved in regulating or modulating the timing and/or rate of transcription of a nucleic acid of the invention. For example, a promoter can be a *cis*-acting transcriptional control element, including an enhancer, a promoter, a transcription terminator, an origin of replication, a chromosomal integration sequence, 5' and 3' untranslated regions, or an intronic sequence, which are involved in transcriptional regulation. These *cis*-acting sequences typically interact with proteins or other biomolecules to carry out (turn on/off, regulate, modulate, etc.) transcription.

As used herein, "recombinant" refers to a polynucleotide synthesized or otherwise manipulated *in vitro* (e.g., "recombinant polynucleotide"), to methods of using recombinant polynucleotides to produce gene products in cells or other biological systems, or to a polypeptide ("recombinant protein") encoded by a recombinant polynucleotide. "Recombinant means" also encompass the ligation of nucleic acids having various coding regions or domains or promoter sequences from different sources into an expression cassette or vector for expression of, e.g., inducible or constitutive expression of polypeptide coding sequences in the vectors of invention.

As used herein, the "sequence" of a nucleic acid or gene refers to the order of nucleotides in the polynucleotide, including either or both strands (sense and antisense) of a double-stranded DNA molecule, *e.g.*, the sequence of both the coding strand and its complement, or of a single-stranded nucleic acid molecule (sense or antisense). For example, in alternative embodiments, promoters drive the transcription of sense and/or antisense polynucleotide sequences of the invention, as exemplified by SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:5, and SEQ ID NO:7.

The terms "identical" or percent "sequence identity," in the context of two or more nucleic acids or polypeptide sequences, refer to two or more sequences or subsequences that are

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the same or have a specified percentage of nucleotides (or amino acid residues) that are the same, when compared and aligned for maximum correspondence over a comparison window, as measured using one of the following sequence comparison algorithms or by manual alignment and visual inspection. This definition also refers to the complement (antisense strand) of a sequence. For example, in alternative embodiments, nucleic acids within the scope of the invention include those with a nucleotide sequence identity that is at least about 60%, at least about 65%, at least about 70%, at least about 75%, at least about 80%, at least about 85%, at least about 90%, and at least about 95% of the exemplary sequences set forth in SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:5, and SEQ ID NO:7; or, SEQ ID NO:9 through SEQ ID NO:14.

In alternative embodiments, polypeptides within the scope of the invention include those with an amino acid sequence identity that is at least about 60%, at least about 65%, at least about 70%, at least about 75%, at least about 80%, at least about 85%, at least about 90%, and at least about 95% of the exemplary sequences set forth in SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, and SEQ ID NO:8.

Two sequences with these levels of identity are "substantially identical" and within the scope of the invention. Thus, if a nucleic acid sequence has the requisite sequence identity to SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:5, or SEQ ID NO:7, or, SEQ ID NO:9 through SEQ ID NO:14, or a subsequence thereof, it also is a polynucleotide sequence within the scope of the invention. If a polynucleotide sequence has the requisite sequence identity to SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, and SEQ ID NO:8, or a subsequence thereof, it also is a polypeptide within the scope of the invention.

The percent identity can exists over a region of the sequence that is at least about 25 nucleotides or amino acid residues in length, or, alternatively, over a region that is at least about 50 to 100 nucleotides or amino acids in length. Parameters (including, e.g., window sizes, gap penalties and the like) to be used in calculating "percent sequence identities" between two nucleic acids or polypeptides to identify and determine whether one is within the scope of the invention are described in detail, below.

The phrase "selectively (or specifically) hybridizes to" refers to the binding, duplexing, or hybridizing of a molecule to a particular nucleotide sequence under stringent hybridization conditions when that sequence is present in a complex mixture (e.g., total cellular or library DNA or RNA), wherein the particular nucleotide sequence is detected at least at about 10 times

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background. In one embodiment, a nucleic acid can be determined to be within the scope of the invention (e.g., is substantially identical to SEQ ID NO:1, 3, 5, or 7, or, SEQ ID NO:9 through SEQ ID NO:14) by its ability to hybridize under stringent conditions to a nucleic acid otherwise determined to be within the scope of the invention (such as the exemplary sequences described herein).

The phrase "stringent hybridization conditions" refers to conditions under which a probe will primarily hybridize to its target subsequence, typically in a complex mixture of nucleic acid, but to no other sequences in significant amounts, is described in detail below. A positive signal (e.g., identification of a nucleic acid of the invention) is about 10 times background hybridization.

Nucleic Acids, Vectors and Primer Pairs

This invention provides novel nucleic acids for use in the diagnosis and treatment of Giant Cell Arthritis (GCA) and means to make and express those nucleic acids. As the genes and vectors of the invention can be made and expressed in vitro or in vivo, the invention provides for a variety of means of making and expressing these genes and vectors. One of skill will recognize that desired phenotypes associated with altered gene activity can be obtained by modulating the expression or activity of the genes and nucleic acids (e.g., promoters) within the vectors of the invention. Any of the known methods described for increasing or decreasing expression or activity can be used for this invention. The invention can be practiced in conjunction with any method or protocol known in the art, which are well described in the scientific and patent literature.

General Techniques

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The nucleic acid sequences of the invention and other nucleic acids used to practice this invention, whether RNA, cDNA, genomic DNA, vectors, viruses or hybrids thereof, may be isolated from a variety of sources, genetically engineered, amplified, and/or expressed recombinantly. Any recombinant expression system can be used, including, in addition to bacterial cells, e.g., mammalian, yeast, insect or plant cell expression systems.

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Alternatively, these nucleic acids can be synthesized *in vitro* by well-known chemical synthesis techniques, as described in, e.g., Carruthers (1982) Cold Spring Harbor Symp. Quant. Biol. 47:411-418; Adams (1983) J. Am. Chem. Soc. 105:661; Belousov (1997) Nucleic Acids Res. 25:3440-3444; Frenkel (1995) Free Radic. Biol. Med. 19:373-380; Blommers (1994) Biochemistry 33:7886-7896; Narang (1979) Meth. Enzymol. 68:90; Brown (1979) Meth. Enzymol. 68:109; Beaucage (1981) Tetra. Lett. 22:1859; U.S. Patent No. 4,458,066. Double stranded DNA fragments may then be obtained either by synthesizing the complementary strand and annealing the strands together under appropriate conditions, or by adding the complementary strand using DNA polymerase with an appropriate primer sequence.

Techniques for the manipulation of nucleic acids, such as, e.g., generating mutations in sequences, subcloning, labeling probes, sequencing, hybridization and the like are well described in the scientific and patent literature, see, e.g., Sambrook, ed., Molecular Cloning: A Laboratory Manual (2nd ed.), Vols. 1-3, Cold Spring Harbor Laboratory, (1989); Current Protocols in Molecular Biology, Ausubel, ed. John Wiley & Sons, Inc., New York (1997); Laboratory Techniques in Biochemistry and Molecular Biology: Hybridization With Nucleic Acid Probes, Part I. Theory and Nucleic Acid Preparation, Tijssen, ed. Elsevier, N.Y. (1993).

Nucleic acids, vectors, capsids, polypeptides, and the like can be analyzed and quantified by any of a number of general means well known to those of skill in the art. These include, e.g., analytical biochemical methods such as NMR, spectrophotometry, radiography, electrophoresis, capillary electrophoresis, high performance liquid chromatography (HPLC), thin layer chromatography (TLC), and hyperdiffusion chromatography, various immunological methods, e.g. fluid or gel precipitin reactions, immunodiffusion, immuno-electrophoresis, radioimmunoassays (RIAs), enzyme-linked immunosorbent assays (ELISAs), immunofluorescent assays, Southern analysis, Northern analysis, dot-blot analysis, gel electrophoresis (e.g., SDS-PAGE), RT-PCR, quantitative PCR, other nucleic acid or target or signal amplification methods, radiolabeling, scintillation counting, and affinity chromatography.

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Classical Diagnosis, Biopsies and Microdissections of Arteritis Lesions

The invention provides compositions and methods for isolating novel GCA-associated nucleic acids and polypeptides within the scope of the invention. A preferred source of such GCA-associated tissue is from arteritis lesions. Temporal artery segments biopsied from the same individual can yield both normal and arteritis tissue samples. Histopathological analysis can confirm both "normal" and arteritis samples.

In addition to the diagnostic methods provided by the invention, GCA diagnosis can be complemented by histopathologic analysis. The histopathology of a GCA lesion typically shows chronic granulomatous inflammation, often with abnormally large multinucleated cells (giant cells), and destruction of the internal elastic lamina of the artery. Chronic inflammation is sometimes confined to the different branches of the heart's main artery (aorta) and any large arteries can become inflamed. The temporal arteries of the head are most frequently affected (temporal arteritis). In rare cases, veins may also be affected by giant cell arteritis.

Examination of the cellular inflammatory infiltrate in GCA lesions primarily reveals macrophages and CD4+ T cells. Immunohistochemistry demonstrates the diffuse presence of IL-6 or IL-1 β expressing CD68+ macrophages. CD68+ cells expressing 72kD type IV collagenase and the inducible nitric oxide synthase (iNOS) are found in the intima and intimamedia of the artery. It is interesting to note that B cells are infrequent in GCA lesions. Lesional B cells and plasma cells are identified in GCA lesions by expression of CD20 and light chain, respectively. These cells can be found in GCA-affected adventitia, both scattered and located in perivascular clusters, in the same tissue microenvironment as the antigenically-activated T cells. These observations suggest that the artery is the site for antigenic encounter and suggest that GCA is mediated by antigen-specific pathogenic T cells.

Clinical symptoms can also help diagnose GCA. The symptoms of GCA may include fatigue, malaise, weight loss, stiffness, muscle pain, fever, and/or headaches. Generalized muscle pain, claudication of the jaw or tongue, or localized scalp or temporal pain, swelling, and tenderness are also common manifestations of GCA. Headache, usually temporal, is present in about 66% of cases. Headache usually begins early in the course of the disease and may be the presenting symptom. The pain is severe and localized to the temple. Scalp tenderness is common. The local vessels are thickened, and tender. Occasionally they are visible. Visual disturbances have been seen in about 25 to 50% of cases. Blindness is the most serious and

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irreversible feature of GCA, where it can be sudden, painless and permanent. Involvement of both eyes can occur. Blindness may be the initial clinical presentation of GCA.

Neuro-ophthalmic complications include arteritic anterior ischemic optic neuropathy, posterior optic neuropathy, choroidal ischemia, diplopia, retinal arterial occlusions, and ocular ischemic syndromes and occur in up to 70% of GCA. Notably, it is estimated that 15 to 20% of patients with GCA suffer permanent and potentially bilateral visual loss from ischemic infarction of the optic nerve.

Currently used methods to diagnosis GCA rely on an extracranial arterial biopsy that demonstrates a necrotizing vasculitis with mononuclear cell infiltration or granulomatous inflammation. Pathologic evidence of GCA in arterial biopsies can confirm the diagnosis but, because the arterial involvement is discontinuous and may show skip areas, a negative biopsy does not rule out GCA. Thus, in the absence of pathologic confirmation, diagnosis would rely on the presence of a constellation of non-specific signs or symptoms. However, the invention provides additional, more definitive diagnostic (and treatment) procedures.

One exemplary means to biopsy or isolate GCA lesions is by dissection with lasercapture microdissection (LCM). Either treshly biopsied or archival pathology specimens of GCA-positive arteries from both histopathologically involved and uninvolved areas. Because the vasculitis of GCA occurs in an irregular, or discontinuous pattern (the co-called "skip lesion"), isolation of one artery (or vein, if appropriate) sample can yield both involved and uninvolved tissue samples. Retrieval of selected cells is achieved by activation of a transfer film placed in contact with a tissue section, by a laser beam (30 or 60 micron diameter) that is focused on a selected area of tissue using an inverted microscope. In LCM, a thermoplastic polymer coating (e.g., ethylene vinyl acetate) attached to a rigid support is placed in contact with a tissue section. The EVA polymer over microscopically selected cell clusters is precisely activated by a nearinfrared laser pulse and bonds to the targeted area\ Removal of the EVA and its support from the tissue section procures the selected cell aggregates for molecular analysis. A computer-controlled arm can precisely position a 40-micron-wide strip of a cylindrical EVA surface onto a sample with a light contact force. Techniques of laser-capture microdissection are known in the art, e.g., the PixCell laser capture microdissection (LCM) system, see, e.g., Kohda (2000) Kidney Int. 57:321-331; Goldsworthy (1999) Mol. Carcinog. 25:86(91; Banks (1999) Electrophoresis

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20:689-700; Emmert-Buck (1996) Science 274:998-1001; U.S. Patent Nos. 5,985,085; 5,859,699.

Hybridization for Identifying Nucleic Acids of the Invention

Nucleic acids within the scope of the invention include isolated or recombinant nucleic acids which specifically hybridizes to an exemplary nucleic acid of the invention. Stringent conditions are sequence-dependent and will be different in different circumstances. Longer sequences hybridize specifically at higher temperatures. An extensive guide to the hybridization of nucleic acids is found in Tijssen, Techniques in Biochemistry and Molecular Biology--Hybridization with Nucleic Probes, "Overview of principles of hybridization and the strategy of nucleic acid assays" (1993). Generally, stringent conditions are selected to be about 5 to 10°C lower than the thermal melting point (Tm) for the specific sequence at a defined ionic strength and pH. The Tm is the temperature (under defined ionic strength, pH, and nucleic acid concentration) at which 50% of the probes complementary to the target hybridize to the target sequence at equilibrium (as the target sequences are present in excess, at Tm, 50% of the probes are occupied at equilibrium). Stringent conditions will be those in which the salt concentration is less than about 1.0 M sodium ion, typically about 0.01 to 1.0 M sodium ion concentration (or other salts) at pH 7.0 to 8.3 and the temperature is at least about 30°C for short probes (e.g., 10 to 50 nucleotides) and at least about 60°C for long probes (e.g., greater than 50 nucleotides). Stringent conditions may also be achieved with the addition of destabilizing agents such as formamide.

For selective or specific hybridization, a positive signal (e.g., identification of a nucleic acid of the invention) is about 10 times background hybridization. "Stringent" hybridization conditions that are used to identify substantially identical nucleic acids within the scope of the invention include hybridization in a buffer comprising 50% formamide, 5x SSC, and 1% SDS at 42°C, or hybridization in a buffer comprising 5x SSC and 1% SDS at 65°C, both with a wash of 0.2x SSC and 0.1% SDS at 65°C. Exemplary "moderately stringent hybridization conditions" include a hybridization in a buffer of 40% formamide, 1 M NaCl, and 1% SDS at 37°C, and a wash in 1X SSC at 45°C. Those of ordinary skill will readily recognize that alternative but

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comparable hybridization and wash conditions can be utilized to provide conditions of similar stringency.

Nucleic acids which do not hybridize to each other under moderately stringent or stringent hybridization conditions are still substantially identical if the polypeptides which they encode are substantially identical. This may occur, *e.g.*, when a copy of a nucleic acid is created using the maximum codon degeneracy permitted by the genetic code, as discussed herein (see discussion on "conservative substitutions").

However, the selection of a hybridization format is not critical - it is the stringency of the wash conditions that set forth the conditions which determine whether a nucleic acid is within the scope of the invention. Wash conditions used to identify nucleic acids within the scope of the invention include, e.g.: a salt concentration of about 0.02 molar at pH 7 and a temperature of at least about 50°C or about 55°C to about 60°C; or, a salt concentration of about 0.15 M NaCl at 72°C for about 15 minutes; or, a salt concentration of about 0.2X SSC at a temperature of at least about 50°C or about 55°C to about 60°C for about 15 to about 20 minutes; or, the hybridization complex is washed twice with a solution with a salt concentration of about 2X SSC containing 0.1% SDS at room temperature for 15 minutes and then washed twice by 0.1X SSC containing 0.1% SDS at 68°C for 15 minutes; or, equivalent conditions. See Sambrook, Tijssen and Ausubel for a description of SSC buffer and equivalent conditions.

Amplification of Nucleic Acids

The invention provides oligonucleotide primers that can amplify nucleic acid encoding GCA-associated nucleic acids. Thus, nucleic acids of the invention can be, e.g., subcloned or measured quantitatively using amplification techniques. Amplification can also be used to identify complementary GCA-associated nucleic acids in tissue biopsies (e.g., from arteritis lesions) or fluid (e.g., serum, whole blood, urine) samples to aid in the diagnosis or treatment of GCA (see below).

Using the exemplary degenerate primer pair sequences of the invention (see below), the skilled artisan can select and design suitable oligonucleotide amplification primers.

Amplification methods are also well known in the art, and include, e.g., polymerase chain reaction, PCR (PCR PROTOCOLS, A GUIDE TO METHODS AND APPLICATIONS, ed.

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Innis, Academic Press, N.Y. (1990) and PCR STRATEGIES (1995), ed. Innis, Academic Press, Inc., N.Y., ligase chain reaction (LCR) (see, e.g., Wu (1989) Genomics 4:560; Landegren (1988) Science 241:1077; Barringer (1990) Gene 89:117); transcription amplification (see, e.g., Kwoh (1989) Proc. Natl. Acad. Sci. USA 86:1173); and, self-sustained sequence replication (see, e.g., Guatelli (1990) Proc. Natl. Acad. Sci. USA 87:1874); Q Beta replicase amplification (see, e.g., Smith (1997) J. Clin. Microbiol. 35:1477-1491), automated Q-beta replicase amplification assay (see, e.g., Burg (1996) Mol. Cell. Probes 10:257-271) and other RNA polymerase mediated techniques (e.g., NASBA, Cangene, Mississauga, Ontario); see also Berger (1987) Methods Enzymol. 152:307-316; Sambrook; Ausubel; U.S. Patent Nos. 4,683,195 and 4,683,202; Sooknanan (1995) Biotechnology 13:563-564. A continuous amplification reaction method is described by, e.g., U.S. Patent No. 5,981,179.

Once amplified, the libraries can be cloned, if desired, into any of a variety of vectors using routine molecular biological methods; methods for cloning *in vitro* amplified nucleic acids are described, *e.g.*, U.S. Pat. No. 5,426,039. To facilitate cloning of amplified sequences, restriction enzyme sites can be "built into" the PCR primer pair.

Degenerate Primer Design

In alternative embodiments, primer pairs of the invention are also designed to selectively amplify new nucleic acid sequences found only in arteritis lesions and not in normal vasculature or other normal tissues. To amplify GCA-associated nucleic acids within the scope of the invention, degenerate primer pairs are designed based on the exemplary nucleic acids of the invention, e.g., SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:5, and SEQ ID NO:7; or, SEQ ID NO:9 through SEQ ID NO:14.

Paradigms to design degenerate primer pairs are well known in the art. For example, a COnsensus-DEgenerate Hybrid Oligonucleotide Primer (CODEHOP) strategy computer program is accessible as http://blocks.fhcrc.org/codehop.html, and is directly linked from the BlockMaker multiple sequence alignment site for hybrid primer prediction beginning with a set of related protein sequences, (see, e.g., Rose (1998) Nucleic Acids Res. 26:1628-1635; Singh (1998) Biotechniques 24:318-319).

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Means to synthesize oligonucleotide primer pairs, including degenerate primer pairs, are well known in the art. "Natural" base pairs or synthetic base pairs can be used. For example, use of artificial nucleobases offers a versatile approach to manipulate primer sequence and generate a more complex mixture of amplification products. Various families of artificial nucleobases are capable of assuming multiple hydrogen bonding orientations through internal bond rotations to provide a means for degenerate molecular recognition. Incorporation of these analogs into a single position of a PCR primer allows for generation of a complex library of amplification products. See, e.g., Hoops (1997) Nucleic Acids Res. 25:4866-4871. Nonpolar molecules can also be used to mimic the shape of natural DNA bases. A non-hydrogen-bonding shape mimic for adenine can replicate efficiently and selectively against a nonpolar shape mimic for thymine (see, e.g., Morales (1998) Nat. Struct. Biol. 5:950-954). For example, two degenerate bases can be the pyrimidine base 6H, 8H-3,4-dihydropyrimido[4,5-c][1,2]oxazin-7one or the purine base N6-methoxy-2,6-diaminopurine (see, e.g., Hill (1998) Proc. Natl. Acad. Sci. USA 95:4258-4263). Exemplary primers of the invention can also incorporate the nucleobase analog 5'-Dimethoxytrityl-N-benzoyl-2'-deoxy-Cytidine,3'-[(2-cyanoethyl)-(N,Ndiisopropyl)]-phosphoramidite. This pyrimidine analog hydrogen bonds with purines, including A and G residues.

Generating and Isolating Novel Nucleic Acids Derived from GCA Lesions

The invention provides compositions and methods for identifying the GCA-associated nucleic acids of the invention in biopsies and tissue samples for diagnostic purposes and to identify new GCA associated nucleic acids and polypeptides. In addition using degenerate primers, as described above, subtractive hybridization of nucleic acids from normal tissue against nucleic acids from arteritis tissue samples (e.g., from biopsies) can be used to isolate GCA-associated nucleic acids within the scope of the invention.

The isolation of nucleic acids may be accomplished by a number of techniques, all well known in the art. For instance, oligonucleotide probes (e.g., PCR primers or hybridization probes) based on the sequences disclosed herein can be used to identify desired nucleic acids in a cDNA or a genomic DNA library. To construct genomic libraries, large segments of genomic DNA are generated by random fragmentation, e.g. using restriction endonucleases, and are

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ligated with vector DNA to form concatemers that can be packaged into the appropriate vector. To prepare a cDNA library, mRNA is isolated (e.g., from a GCA lesion biopsy), and a cDNA library containing sequences encoding GCA-associated polypeptides is prepared from the mRNA. The cDNA or genomic library can then be screened using a probe based upon the sequence of a cloned gene disclosed herein. Probes may be used to hybridize with genomic DNA or cDNA sequences to isolate homologous genes (new GCA associated sequences) or to diagnose GCA. As discussed herein, defined stringent hybridization conditions can be used to identify nucleic acid sequences within the scope of the invention.

Alternatively, antibodies raised against a GCA-associated polypeptide (see below) can be used to screen a cDNA expression library. Alternatively, the nucleic acids of interest can be amplified from nucleic acid samples using amplification techniques.

As noted above, subtractive hybridization of nucleic acid of normal from GCA-involved tissue is another exemplary means of isolating GCA-associated nucleic acid. Subtractive hybridization techniques are well known in the art, e.g., as micro-genomic representational difference analysis (RDA); see, e.g., Lisitsyn (1993) Science 259:946-951; Michiels (1998) Nucleic Acids Res. 26:3608-3610; Wan (1996) Nat. Biotechnol. 14:1685-1691; U.S. Patent Nos. 6,013,437; 5,958,738; 5,935,788; 5,882,874; 5,525,471.

Cloning and construction of expression vectors

The invention provides libraries of expression vectors comprising the GCA polypeptide-encoding sequences of the invention. These nucleic acids may be introduced into a genome or into the cytoplasm or a nucleus of a cell and expressed by a variety of conventional techniques, well described in the scientific and patent literature. See, e.g., Roberts (1987) Nature 328:731; Berger (1987) supra; Schneider (1995) Protein Expr. Purif. 6435:10; Sambrook, Tijssen or Ausubel. Product information from manufacturers of biological reagents and experimental equipment also provide information regarding known biological methods. The vectors can be isolated from natural sources, obtained from such sources as ATCC or GenBank libraries, or prepared by synthetic or recombinant methods.

The nucleic acids of the invention can be expressed in expression cassettes, vectors or viruses which are stably or transiently expressed in cells (e.g., episomal expression systems).

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Selection markers can be incorporated into expression cassettes and vectors to confer a selectable phenotype on transformed cells and sequences. For example, selection markers can code for episomal maintenance and replication such that integration into the host genome is not required. For example, the marker may encode antibiotic resistance (e.g., chloramphenicol, kanamycin, G418, bleomycin, hygromycin) or herbicide resistance (e.g., chlorosulfuron or Basta) to permit selection of those cells transformed with the desired DNA sequences (see, e.g., Blondelet-Rouault (1997) Gene 190:315-317; Aubrecht (1997) J. Pharmacol. Exp. Ther. 281:992-997). Because selectable marker genes conferring resistance to substrates like neomycin or hygromycin can only be utilized in tissue culture, chemoresistance genes can be used as selectable markers *in vitro* and *in vivo*.

Alignment Analysis of Gene Sequences

The nucleic acid sequences of the invention include genes and gene products identified and characterized by analysis using the exemplary nucleic acid and protein sequences of the invention, including, e.g., the exemplary SEQ ID NO:1 and SEQ ID NO:2; SEQ ID NO:3 and SEQ ID NO:4; SEQ ID NO:5 and SEQ ID NO:6; SEQ ID NO:7 and SEQ ID NO:8, respectively. For sequence comparison, typically one sequence acts as a reference sequence, to which test sequences are compared. When using a sequence comparison algorithm, test and reference sequences are entered into a computer, subsequence coordinates are designated, if necessary, and sequence algorithm program parameters are designated. Default program parameters are used unless alternative parameters are designated herein. The sequence comparison algorithm then calculates the percent sequence identity for the test sequence(s) relative to the reference sequence, based on the designated or default program parameters. A "comparison window", as used herein, includes reference to a segment of any one of the number of contiguous positions selected from the group consisting of from 25 to 600, usually about 50 to about 200, more usually about 100 to about 150 in which a sequence may be compared to a reference sequence of the same number of contiguous positions after the two sequences are optimally aligned.

Methods of alignment of sequences for comparison are well-known in the art. Optimal alignment of sequences for comparison can be conducted, e.g., by the local homology algorithm of Smith & Waterman, Adv. Appl. Math. 2:482 (1981), by the homology alignment algorithm of

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Needleman & Wunsch, J. Mol. Biol. 48:443 (1970), by the search for similarity method of Pearson & Lipman, Proc. Natl. Acad. Sci. USA 85:2444 (1988), by computerized implementations of these algorithms (CLUSTAL, GAP, BESTFIT, FASTA, and TFASTA in the Wisconsin Genetics Software Package, Genetics Computer Group, 575 Science Dr., Madison, WI), or by manual alignment and visual inspection.

In a preferred embodiment, CLUSTAL algorithm is used, particularly, the CLUSTAL W program, see, *e.g.*, Thompson (1994) Nuc. Acids Res. 22:4673-4680; Higgins (1996) Methods Enzymol 266:383-402. Variations can also be used, such as CLUSTAL X, see Jeanmougin (1998) Trends Biochem Sci 23:403-405; Thompson (1997) Nucleic Acids Res 25:4876-4882 CLUSTAL is a particularly preferred program for determining if sequences are so substantially identical they are within the scope of the invention because, if a comparison set consists of enough closely related sequences so that the first alignments are accurate, then CLUSTAL W will usually find an alignment that is very close to ideal. In one embodiment, the CLUSTAL W program described by Thompson (1994) supra, is used with the following parameters: K tuple (word) size: 1, window size: 5, scoring method: percentage, number of top diagonals: 5, gap penalty: 3, to determine whether a nucleic acid has sufficient sequence identity to an exemplary nucleic acid (SEQ ID NO:1, 3, 5, or 7) to be with the scope of the invention.

Another algorithm is PILEUP, which can be used to determine whether a nucleic acid has sufficient sequence identity to SEQ ID NO:1, 3, 5, or 7, or SEQ ID NO:9 through SEQ ID NO:14, to be with the scope of the invention. This program creates a multiple sequence alignment from a group of related sequences using progressive, pairwise alignments to show relationship and percent sequence identity. It also plots a tree or dendogram showing the clustering relationships used to create the alignment. PILEUP uses a simplification of the progressive alignment method of Feng & Doolittle, J. Mol. Evol. 35:351-360 (1987). The method used is similar to the method described by Higgins & Sharp, CABIOS 5:151-153 (1989). Using PILEUP, a reference sequence (e.g., an exemplary GCA-associated sequence of the invention) is compared to another sequence to determine the percent sequence identity relationship (i.e., that the second sequence is substantially identical and within the scope of the invention) using the following parameters: default gap weight (3.00), default gap length weight (0.10), and weighted end gaps. In one embodiment, PILEUP obtained from the GCG sequence analysis software package, e.g., version 7.0 (Devereaux(1984) Nuc. Acids Res. 12:387-395),

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using the parameters described therein, is used to identify nucleic acids within the scope of the invention.

Another example of an algorithm that is suitable for determining percent sequence identity (i.e., substantial similarity or identity) in this invention is the BLAST algorithm, which is described in Altschul (1990) J. Mol. Biol. 215:403-410. Software for performing BLAST analyses is publicly available through the National Center for Biotechnology Information (http://www.ncbi.nlm.nih.gov/). This algorithm involves first identifying high scoring sequence pairs (HSPs) by identifying short words of length W in the query sequence, which either match or satisfy some positive-valued threshold score T when aligned with a word of the same length in a database sequence. T is referred to as the neighborhood word score threshold (Altschul (1990) supra). These initial neighborhood word hits act as seeds for initiating searches to find longer HSPs containing them. The word hits are then extended in both directions along each sequence for as far as the cumulative alignment score can be increased. Cumulative scores are calculated using, for nucleotide sequences, the parameters M (reward score for a pair of matching residues; always > 0) and N (penalty score for mismatching residues, always < 0). For amino acid sequences, a scoring matrix is used to calculate the cumulative score. Extension of the word hits in each direction are halted when: the cumulative alignment score falls off by the quantity X from its maximum achieved value; the cumulative score goes to zero or below, due to the accumulation of one or more negative-scoring residue alignments; or the end of either sequence is reached. The BLAST algorithm parameters W, T, and X determine the sensitivity and speed of the alignment. In one embodiment, to determine if a nucleic acid sequence is within the scope of the invention, the BLASTN program (for nucleotide sequences) is used incorporating as defaults a wordlength (W) of 11, an expectation (E) of 10, M=5, N=4, and a comparison of both strands. For amino acid sequences, the BLASTP program uses as default parameters a wordlength (W) of 3, an expectation (E) of 10, and the BLOSUM62 scoring matrix (see, e.g., Henikoff (1989) Proc. Natl. Acad. Sci. USA 89:109 (5).

Exemplary GCA-Associated Nucleic Acids and Polypeptides

The invention provides novel GCA-associated nucleic acids and polypeptides. All possible polypeptides encoded by these nucleic acids are within the scope of the invention,

including alterative in-frame ORFs and ORFs read from either strand (e.g., including the complementary strand). For example:

>GCA 1a = 1c.T7 cloned sequence of GCA1 and deduced protein sequence 5 1 - GATCCCCGCTTTCGCGGGGATGACAGCGGTACTCAATTCACGCGCAGCGATGCCAGCGAA - 60 61 - CTAAACGGAGGATCTCACGAACATCCGCTCCAACCCCGACACCACGCTCCCCGCCGTCAC - 120 L P A V T 10 121 - GACAGGCTCGCTCCTCCCCCCCAAGTTCTTTGCAATCCCTGAGGCCGCCCCGACAT - 180 T G S L P S S R K F F A I P E A A P D I 181 - CCGCGTTCCCTTGCGCGAGATCATCCTGTCCGAGGGCGCCGAGCCGAACCTGCCGGT - 240 - R V P L R E I I L S E G A G E P N L P V 15 241 - CTATGACACCTCGGGCCCTACACCGATCCGGCCGTGACGATCGACGTCAACAGCGGCCT - 300 $\begin{smallmatrix} Y & D & T & S & G & P & Y & T & D & P & A & V & T & I & D & V & N \\ \end{smallmatrix}$ 301 - GCCGCGCAATCGCCTCGCCTGGGTCAAGGAACGCGGCGGCGTCGAGGAATATCAGGCCGC - 360 - PRNRLAWVKERGGVEEYQAA 361 - ACCATCAAGCCGGAGGACAACGGCAATGTCGGCGCCATCCCACGCCGCCAAGGCGTTCACC - 420 - PSSRRTT SEQ ID NO:2 421 - GGCACCACAAGCCGCTGCGCGGCTCGACGGCACAAGATCACCCACTCGAGTTCGCCGCGC - 480 481 - CGGCATTATACCAAGGAGATGATCTACGTCGCCGAGCGTGAGAATCTTGGCGCAAGCAGC - 540 **30** 541 - AGCTGAGCGCCGAGGCCGGCTGCCGACGGAAGAGTTTTGGCGCCGCGGTGCCGGCTTA - 600 601 - TTACGCCGGAATTTGTCGCAAGAGATCGCGCGGCGGCCATTATTTCCTTTAAAATTAACA - 660 **D** SEQ ID NO:1 661 - TTGCCGAGCTTGAACCGATGAA - 682 35 GCA1a: full length clone GATCCCCGCTTTCGCGGGGATGACAGCGGTACTCAATTCACGCGCAGCGA -50TGCCAGCGAACTAAACGGAGGATCTCACGAACATCCGCTCCAACCCCGAC -100ACCACGCTCCCCGCCGTCACGACAGGCTCGCTGCCCTCCTCGCGCAAGTT -150CTTTGCAATCCCTGAGGCCGCGCCCGACATCCGCGTTCCCTTGCGCGAGA -200 40 -250TCATCCTGTCCGAGGGCGCGGCGAGCCGAACCTGCCGGTCTATGACACC TCGGGCCCTACACCGATCCGGCCGTGACGATCGACGTCAACAGCGGCCT -300GCCGCGCAATCGCCTCGCCTGGGTCAAGGAACGCGGCGGCGTCGAGGAAT -350ATCANGGCCGCACCATCAAGCCGGAGGACAACGGCAATGTCGGCGCATCC -400-450 CACGCCGCCAAGGCGTTCACCGNGCACCACAAGCCGCTGCGCGGNCTCGA 45 -500 CGGCACAAGATCACCCACTCGAGTTCGCCGCCGCCGCATTATACCAAGGA GATGATCTACGTCGCCGAGCGTGAGAATCTTGGNCGCAAGCAGCAGCTNG -550-600 AGCGCGCCGANGGCCGGCTNGCCGACGGNAAGAGTTTTGGCGCCGCGGTG -700 CCGGNCTTNATTACGCCGGAATTTGTNCGCAANGAGATCGNCGCGGNCGN GCCATTATTTCCTTTNAAAATTAANCATTGCCGAGCTTGAACCGATGAAN N -701 50

Cloned fragment of GCA la used in GCA la-GST fusion protein

	1 - CTCCCCGCCGTCACGACAGGCTCGCTGCCCTCCTCGCGCAAGTTCTTTGCAATCCCTGAG - 60	
		_
_	61 - GCCGCGCCGACATCCGCGTTCCCTTGCGCGAGATCATCCTGTCCGAGGGCGCCGGCGAG - 12	0
5		
	121 - CCGAACCTGCCGGTCTATGACACCTCGGGCCCCTACACCGATCCGGCCGTGACGATCGAC - 16	U
١.		
	181 - GTCAACAGCGGCCTGCCGCGCGCGTCGAGGCGCGCGCGTCGAG - 24	٠
10	_ v n s g l P R N R L A w v k E k G G v 2	
	241 - GAATATCAGGCCGCACCATCAAGCCGGAGGACAACGGC - 278	
	- E Y Q A A P S S R R T T SEQ ID NO:2	
	>GCA 1b = 1cT3 cloned sequence of GCA1 and deduced protein sequence	
45		
15	1 - ACTCTCCAGCCTCTCACCGAGGATGAAGTCGGCTCGTGAAGTGGTTGCGGTCGGGGGCAA - 60	
	- L S S L S P R M K S A R E V V A V G G K	
	61 - AACCCGGGACGAGCTGGCCTTCCTGCCGGCCGCCCTCGAAATTGTCGAGACGCCGCCATC - 120	
20	- TRDELAFLPAALEIVETPPS	
georg,	121 - TCCCACCGCGAGACTCACGGCCGCCTTGCTTGCTTGCTTG	
25	181 - GGCGGGTCTCGGCAGGATCGACATCGTTGCTTCTGCATCCAGAAAGATCGTGCCGGGCGA - 240	
<i>ද</i> ු	- A G L G R I D I V A S A S R K I V P G D	
	n	
	241 - CCGTGTAAAGCTGGTTCAGCCGCTCGAGGTCGGCGTGGTGCGGGCCACTCATGTCCGCGA - 300	
-	- R V K L V Q P L E V G V V R A T H V R D	
3 <u>Q</u>	- 360	
±.	301 - TGGCCAAACCGTCAAGGCCGGCGAGATTCTGATCGAGCTGGATCCATTCGCGGGTGGTGT - 360	
	- G Q T V K A G E I L I E L D P F A G G V	
	361 - GGATGTTGCGCCCCGTCAGAGGTCCATCACGGTGTCGGCGCCCCACGGATCGCCACACCA - 420	
<u>}.</u>	- D V A T SEQ ID NO:4	
35 .	·	20.
	421 - TCTTGTCGACCTTTCTTCACCGACGAGTCACCGCCGAGTTGCCGATATTGCGTGATCTTA - 48	JU
	481 - TCAGAATGCGGCGATGATCAT - 501 SEQ ID NO:3	
	GCA1b: full length clone -50	
40	ACTOTO ACCOMENCIO CACCO ACCO ACCO ACCO ACCO ACCO ACCO	
	TOCCCCCCANANCCCGGGACGAGCTGGCCTTCCTGCCGGCCGCCGCCGCCGCCGCCGCCGCC	
	λ ΨΨCΨCCACACGCCGCCATCTCCCACCGCGAGACTCACGCCGCGTTGGT	
	TCCTCCCTTCCCCCCCCCCCCCCCCCCCCCCCCCCCCC	
	7/7/T/CATICATION AND AND AND AND AND AND AND AND AND AN	
45	Chuldhin CMCCCLCCACATCACATCACATCACATCACATCACATCAC	
	TCCCCAAACCGTCAAGGCCGGCGAGATTCTGATCGAGCTGGATCCATTCG -550	
	CCCCTCTCTCTCTCTCTCTCCCCCCCCCCCCCCCCCCC	
	CCCCANCGCATCCACCATCTTGTCGACCTNTTCTTOTICGCTTCTTCTTCTTCTTCTTCTTCTTCTTCTTCTTCTTCT	
	GTCACCGCGAGTTGCCGATATAGGANAGATCTTANTCANGITAMTOOO	
50	NCGATGATCAT -511 SEO ID NO:3	
	1	

Cloned fragment of GCA 1b used in GCA 1b-GST fusion

	1																					CAAA	-	60
5	61		L AC											R CCT						G GCC		K ATCT	_	120
Ü	01		T		D									L			v			P				120
	121	-	CC																			GTGG	-	180
	101		P	T		R								A								W CGAC		240
10	191	_			L									TGC.			AAA K			P			_	240
	241	_																				CGAT	_	300
														V						V				
	301	-																				TGTG	-	360
15	361	_	G GA	_					G	Ľ	1	Ţı	Τ	E	Ъ	ט	P	r	А	G	G	V		
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> 20	GCA		4	- c	TOI	nea	se	que	enc	e c	T	CA.	L 4 <u>.</u>	and	ae	eau	cea	pı	OTE	∋ın	se	que	nce	€
	1	_	AC	CGA	CGT	CGA	CTA	TCC	ATG	AAC	GGA	TCC	CTG	CAA	CGA	CAT	CGT	GCG'	TAC	GGC	CTA'	TGAA	_	60
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	121																					CAGC		180
Li II		-	Ι	A	L	P	Ι	D	F	S	A	R	Ι	Α	R	N	T	S	*	SE) I	D NO	: 6	
	181	_	AC	GAG	ACA	GAC	GTC	ACG	GAC	GCG	GTC	GAC.	АСТ	CTG	GCG	GG'	TCC'	TAC'	TAC	GTG(GAG	CGCC	_	240
				0110			010		0	,000	010	00		010			-00	0		0.0	J. 10	0000		
	241	-	TG	ACG	GAT	GAC	CTC	GCC	AAG	CGG	GCC	TGG	GAG	CTG	ATG	SAA	GAG	GTC	GAG	AAG	ATG	GGTG	-	300
	201		CC	7 m.c	ccc	CAC	ccc	አመረ	ccc	יא כיכי	ccm	mcc	aaa	7 7 C	200	~m.c	л пг∕-	CAC	ית תיי	n c m c	200	ACGC		260
35	301	_	GC.	AIG	GCG	CAG	GCG.	ATC	GCG	ACC	GGT	TGG	CCG.	AAG	CGC	JTG	ATC	GAG	CAA	TCT	بالد	ACGC	_	300
	361	_	AΑ	AAG	CAG	GCC	GCG.	ATC	GAT	'CGC	GGC	GAT	CAG	GTG	ATC	GTG	GGC	GTG.	AAC	CGC:	ΓAC	CGGC	_	420
	421	-	CC	GAA	CAG	GAG	CAA	CCG	ATC	GAC	ATT.	ATT	GAG.	ATC	GAC	AAC'	TCG.	ACG	GTT(CGG	GCC'	rccc	-	480
## 40	481	_	AG	АТС	CGG	ጥርጥ	СТС	GCC	GAA	АТС	GAA	AAG	GCG	CGT	CATT	гса	AGG	AAG	ርጥጥር	SAG	rcc	GCGC	_	540
	101		110	0	-	.01	010		 .		0111		000	001	J111 .	. 011	100	110	011	0110		0000		510
	541	-	TC	GGG	GAG	CTG	GCG	TGT	ATT	'GCC	CGC.	ACG	GGT	GAG	GGA/	AAT	CTG	CTG	GCT	GCA	GCG.	ACCG	-	600
	601		7.0	ccc	CCm	ccc	ccc	ccc	CCm	יאככ	CTC	cca	~ 7 ~	א תויביי	חממי	~ n ~	aaa	7 m.c.	000	י ת תרי	יתרטר	nmcc		cci
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	661	_	GC	GAC	CAC	GAG	GCG	GTG	CCG	GAG	GTA	GTG'	TCG	GAC	GTT:	rac	GGC	CGT	GCC'	TAT	GC.	ACGG	_	720
											,													

721 - ATCCGTTCATGGATAGTCGACGTCGGT - 747 SEQ ID NO:5

Cloned fragment of GCA14 used in GCA 14-GST fusion

	1 - GATCCCTGCAACGACATCGTGCGTACGGCCTATGAAGCGCTCGCCGCCGTGCTCGGTGGC - 60
5	61 - ACGCAGTCGCTCCACACCAACTCGTTCGACGAGGCGATCGCGCTGCCGATTGACTTCTCC - 120
9	_ m o g t H m N S F D E A I A L P I D E S
	121 - GCCCGGATCGCCCGCAACACCAGCTGAATCTCCAGCAGCACGAGACAGAC
	_ A R T A R N T S * SEQ ID NO:6
	181 - CGGTCGACACTCTGGCGGGGTCCTACTACGTGGAGCGCCTGACGGATGACCTCGCCAAGC - 240
40	101 - CGGTCGACACTGTGGGGGGGTGTTTTTTTTTTTTTTTTT
10	241 - GGGCCTGGGAGCTGATGGAAGAGGTCGAGAAGATGGGTGGCATGGCGCAGGCGATCGCGA - 300
	241 - GGGCCTGGGAGCTGATGGTAGATGT
	301 - CCGGTTGGCCGAAGCGCCTGATCGAGCAATCTGCGACGCAAAAGCAGGCCGCGATCGAT
	301 - CCGGTTGGCCGAAGCGCTGATCGACCATTGTGGGATCGACCATTGTGGGATCGACCATTGTGGGATCGACCATTGTGGGATCGACCATTGTGGGATCGACCATTGTGGGATCGACCATTGTGGGATCGACCATTGTGGGATCGACCATTGTGGGATCGACCATTGTGGGATCGACCATTGTGGGATCGACCATTGTGGACCATTGTGGACCATTGTGGACCATTGTGGACCATTGTGGACCATTGTACATTGTACATTACATTGTACATTGTACATTGTACATTGTACAT
	361 - GCGGCGATCAGGTGATCGTGGGCGTGAACCGCTACCGGCCCGAACAGGAGCAACCGATCG - 420
15	361 - GCGGCGATCAGGTGATCGTGGGCGTGAACCGGTACCGGCGGATCATCGTGGTGGTGGTGGGGCGTGAACCGGCTACCGGGCGGATCATCGTGGGCGGTGAACCGGCTACCGGGCGGATCATCGTGGGCGGTGAACCGGCTACCGGGCGGATCATCGTGGGCGGTGAACCGGCTACCGGGCGGATCATCGTGGGCGGTGAACCGGCTACCGGGCGGATCATCGTGGGCGGTGAACCGGCTACCGGGCGGATCGTGAACCGGCTACCGGGCGGATCGTACCGGCTACCGGGCGGTACCGGCTACCACACACA
	421 - ACATTATTGAGATCGACAACTCGACGGTTCGGGCCTCCCAGATCCGGTGTCTCGCCGAAA - 480
	421 - ACATTATTGAGATCGACACTCGACGGTTCGGGCCTCCCAGATCGGGTTCTGGGCCTCCCAGATCGGGTTCTGGGCCTCCCAGATCGGGTTCTGGGCCTCCCAGATCGGGTTCTGGGCCTCCCAGATCGGGTTCTGGGCCTCCCAGATCGGGTTCTGGGCCTCCCAGATCGGGTTCTGGGCCTCCCAGATCGGGTTCTGGGCCTCCCAGATCGGGTTCTGGGCCTCCCAGATCGGGTTCTGGGCCTCCCAGATCGGGTTCTGGGCCTCCCAGATCGGGTTCTGGGCCTCCCAGATCGGGTTCTGGGCCTCCCAGATCGGGTTCTGGGCCTCCCAGATCGGGTTCTGGGCCTCCCAGATCGGGTTCTGGGCCTCCCAGATCGGGTTCTGGGCCTCCCAGATCGGGTTCTGGGCCTCCCAGATCCGGTTCTGGGCCTCCCAGATCCGGGTTCTGGGCCTCCCAGATCCGGTTCTGGGCCTCCCAGATCCGGTTCTGGGCCTCCCAGATCCGGGTTCTGGGCCTCCCAGATCCGGTTCTGGGCCTCCCAGATCCGGTTCTGGGCCTCCCAGATCCGGTTCTGGGCCTTCCCAGATCCGGTTCTGGGCCTTCCCAGATCCGGTTCTGGGCCTTCCCAGATCCGGTTCTGGGCCTTCCCAGATCCGGTTCTGGGCCTTCCCAGATCCGGTTCTGGGCCTTCCCAGATCCGGTTCTGGGCCTTCCCAGATCCGGTTCTGGGCCTTCCCAGATCCGGTTCTGGGCCTTCCCAGATCCGGTTCTGGGCCTTCCCAGATCCGGTTCTGGGCCTTCCCAGATCCGGTTCTGGGCCTTCCCAGATCCGGTTCTGGGCCTTCCCAGATCCGGTTCTGGGCCTTCCCAGATCCGGTTCTGGGCCTTCCCAGATCCGGTTCTGGGCCTTCCCAGATCCGGTTCTGGGCCTTCCCAGATCCGGTTCTCCGGGCCTTCCCAGATCCGGTTCTTCTGGGCCTTCCCAGATCCGGGCCTTCCCAGATCCGGTTCTCCAGATCCGGTTCTCTGGGCCTTCCCAGATCTCTCTGGGCCTTCCCAGATCTCTCTGGGCCTTCCCAGATCTCTCTGGGCCTTCCCAGATCTTCTGGGCCTTCCAGATCTCTGGGCCTTCCCAGATCTCCGGGCCTTCCCAGATCTCTCTGGGCCTTCCAGATCTCTGGGCCTTCCCAGATCTCTCTGGGCCTTCCCAGATCTCTCTGGGCCTTCCCAGATCTCTCTGGGCCTTCCCAGATCTCTCTGGGCCTTCCCAGATCTCTCTGGGCCTTCCCAGATCTCTCTGGGCTTCTCTGGGCTTCTTCTGGGCTTCTTCTGGGCTTCTCTGGGCTTCTCTGGGCTTCTT
	CONTRACTOR CONTRACTOR STATE OF A TO NO. 5
	481 - TCGAAAAGGCGCGTGATTCAAGGAAGGTTGAGTCC - 515 SEQ ID NO:5
20	
	CA17 cloned total sequence and deduced protein sequence
	- 60
a	1 - ACTCTCCAGCCTCTCACCGAGGATCATCGACGACATTAAGCAGCTGGCCGACAACGGCGT - 60
	I I D D I K Q L A D N G V
25	120
LL:	61 - GCGCGAATTCACGCTGATCGGACAGAATGTCAACGCCTACCACGGCGGAGGGCCCGACGG - 120
<u>.</u>	- REFT. LIGQNVNAYHGGGPDG
13	100
~ [121 - CCGCGTCTGGCCGCCAAATTGCTGCAGCGACTCGCGGACATTCCAGGCGTCATGCG - 180
30	- R V W P L G K L L Q R L A D I P G V M R
4	
±	181 - GCTGCGTTATTCGATCAGCCATCCGCGCGACGTCGACGACGACGCCTGATCGCCGCGCATCG - 240
	- L R Y S I S H P R D V D D S L I A A H R
la :	
	241 - CGATTTGCCCGGACTGATGCCGTTCGTGCACCTGCCGGTGCAATCGGGGGCCGGACCGGAT - 301
-	- D L P G L M P F V H L P V Q S G A D SEQ ID NO:8
35 111 111	301 - C - 301 SEQ ID NO:7
1000	loned fragment of GCA17 used in GCA17-GST Fusion protein
40	1 - ATCATCGACGACATTAAGCAGCTGGCCGACAACGGCGTGCGCGAATTCACGCTGATCGGA - 60
40	_ T T D D T K O L A D N G V R E F T L l G
	61 - CAGAATGTCAACGCCTACCACGGCGGAGGGCCCGACGGCCGCTCTGGCCGCTCGGCAAA - 120
	- Q N V N A Y H G G G P D G R V W P L G K
	121 - TTGCTGCAGCGACTCGCGGACATTCCAGGCGTCATGCGGCTGCGTTATTCGATCAGCCAT - 180
AE.	
45	- L L Q R L A D I P G V M R L R I S I M 181 - CCGCGCGACGTCGACGCCTGATCGCCGCGCATTGCCCGGACTGATGCCG - 240
	- P R D V D D S L I A A H R D L P G L M P
	241 - TTCGTGCACCTGCCGGTGCAATCGGGGGCCGACCG - 275 SEQ ID NO:7
	- F V H L P V Q S G A D SEQ ID NO:8
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ACTCTCCANCCTCTCACCGAGGATCAGAATAGGTGAAGAGCGAAGACACC -50
GAGAACGTCTGGCCTTGAACGGACAGCGTGCTTGAGTTGGTCGGGGTCAC -100
CACCGGACCCGTGTCCACCGGCGCAGTCACNGTGAAAGCACTTGACCATG -150
ATCCCAGACGGTGCCGTCATCCGCGCGGACCCACANCGTNTCCGCGCCCG -200
ACCGGATTGATAGCTCAGCGACACCAGCTGGCTGCCGTGACGTANTTGT -250
GCTGGTTNGGTGCAAGTGCCACCCCGCTCAAGACAAANTGGCCGCACCTG -300
TGCCCGTGTCCCAAACGTCATATTGGGTCGCAGCACTGTCGAACGGATCA -350
CTGTANGTGCACAGCGACNAANCCGCATANCTCTNGCCGTGGGGCGCAAC -400
GATGTTNNACACCGTCTCAACGGGTACCGTGTCNAGGGGANCATTTACNG -450
GGAAAGCATTCGACCACTCCCCCACACCGTGCCCGCATTTGCGCCGATTC -500
CTTTCATTGATATGTCCACGTCGGTNGGNCTTTAAGCNGGCGGCAACCGC -550
GGTGNAGCTNCACTTTTTGTTCCTTTTATTGANGGTTAATTTGCGCCGTT -600
TGGNCGTAANTNTTTNGAAN -620 SEQ ID NO:9

5'3' GCA2a Frame 1

actctccancctctcaccgaggatcagaataggtgaagagcgaagacaccgagaacgtct TLXPLTEDQNR-RAKTPRTS ggccttgaacggacagcgtgcttgagttggtcggggtcaccaccggacccgtgtccaccg G L E R T A C L S W S G S P P D P C P P gcgcagtcacngtgaaagcacttgaccatgatcccagacggtgccgtcatccgcgcggac AQSX-KHLTMIPDGAVIRAD ccacancgtntccgcgcccgaccggattgatagctcagcgacaccagctgggctgccgtg P X R X R A R P D - - L S D T S W A A V acgtanttgtgctggttnggtgcaagtgccaccccgctcaagacaaantggccgcacctg TXLCWXGASATPLKTXWPHL tgcccgtgtcccaaacgtcatattgggtcgcagcactgtcgaacggatcactgtangtgc P C P K R H I G S Q H C R T D H C X C acagcgacnaanccgcatanctctngccgtggggcgcaacgatgttnnacaccgtctcaa T A X X P H X S X R G A Q R C X T P S Q cgggtaccgtgtcnaggggancatttacngggaaagcattcgaccactcccccacaccgt R V P C X G X H L X G K H S T T P P H R ${\tt gcccgcatttgcccgattcctttcattgatatgtccacgtcggtnggnctttaagcngg}$ ARICADSFH-YVHVGXXLSX cggcaaccgcggtgnagctncactttttgttccttttattganggttaatttgcgcgctt - X L I C A L

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5'3' GCA2a Frame 2

actctccancctctcaccgaggatcagaataggtgaagagcgaagacaccgagaacgtctg L S X L S P R I R I G E E R R H R E R L gccttgaacggacagcgtgcttgagttggtcggggtcaccaccggacccgtgtccaccgg ALNGQRA-VGRGHHRTRVHR cgcagtcacngtgaaagcacttgaccatgatcccagacggtgccgtcatccgcgcggacc R S H X E S T - P - S Q T V P S S A R T cacancgtntccgcgcccgaccggattgatagctcagcgacaccagctgggctgccgtga cgtanttgtgctggttnggtgcaagtgccaccccgctcaagacaaantggccgcacctgt R X C A G X V Q V P P R S R Q X G R T C ${\tt gcccgtgtcccaaacgtcatattgggtcgcagcactgtcgaacggatcactgtangtgca}$ A R V P N V I L G R S T V E R I T V X A cagcgacnaanccgcatanctctngccgtggggcgcaacgatgttnnacaccgtctcaac Q R X X R I X X A V G R N D V X H R L N gggtaccgtgtcnaggggancatttacngggaaagcattcgaccactccccacaccgtg $\tt G \quad Y \quad R \quad V \quad X \quad G \quad X \quad I \quad Y \quad X \quad E \quad S \quad I \quad R \quad P \quad L \quad P \quad H \quad T \quad V$ $\verb|cccgcatttgcgccgattcctttcattgatatgtccacgtcggtnggnctttaagcnggc|$ PAFAPIPFIDMSTS·XXL-XG ${\tt ggcaaccgcggtgnagctncactttttgttccttttattganggttaatttgcgcgcttt}$ G N R G X A X L F V P F I X G - F A R F ggncgtaantntttngaan, SEDIAN

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5'3' GCA2a Frame 3

actctccancctctcaccgaggatcagaataggtgaagagcgaagacaccgagaacgtctgg S P X S H R G S E - V K S E D T E N V W ccttgaacggacagcgtgcttgagttggtcggggtcaccaccggacccgtgtccaccggc P-TDSVLELVGVTTGPVSTG gcagtcacngtgaaagcacttgaccatgatcccagacggtgccgtcatccgcgcggaccc AVXVKALDHDPRRCRHPRGP acancgtntccgcgcccgaccggattgatagctcagcgacaccagctgggctgccgtgac T X X P R P T G L I A Q R H Q L G C R D gtanttgtgctggttnggtgcaagtgccaccccgctcaagacaaantggccgcacctgtg V X V L V X C K C H P A Q D K X A A P V cccgtgtcccaaacgtcatattgggtcgcagcactgtcgaacggatcactgtangtgcac P V S Q T S Y W V A A L S N G S L X V H agcgacnaanccgcatanctctngccgtggggcgcaacgatgttnnacaccgtctcaacgS D X X A X L X P W G A T M X X T V S T ggtaccgtgtcnaggggancatttacngggaaagcattcgaccactccccacaccgtgc G T V X R G X F X G K A F D H S P T P C $\verb|cegcatttgegccgattcctttcattgatatgtccacgtcggtnggnctttaagcnggcg|$ P H L R R F L S L I C P R R X X F K X A gcaaccgcggtgnagctncactttttgttccttttattganggttaatttgcgcgctttg FLLLXVNLRAL A T A V X X H F L gncgtaantntttngaan, х х Х

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3'5' **GCA2a** Frame 1

nttcnaaananttacgnccaaagcgcgcaaattaaccntcaataaaaggaacaaaaagtg X X X X Y X Q S A Q I N X Q - K E Q K V nagctncaccgcggttgccgccngcttaaagnccnaccgacgtggacatatcaatgaaag XXHRGCRXLKXXPTWTYQ-K gaatcggcgcaaatgcgggcacggtgtggggggggtggtcgaatgctttcccngtaaatgn E S A Q M R A R C G G V V E C F P X K X tcccctngacacggtacccgttgagacggtgtnnaacatcgttgcgccccacggcnagag SPXHGTR-DGXXHRCAPRXE ntatgcggnttngtcgctgtgcacntacagtgatccgttcgacagtgctgcgacccaata X C X X V A V H X Q - S V R Q C C D P I tgacgtttgggacacgggcacaggtgcggccantttgtcttgagcggggtggcacttgca - R L G H G H R C G X F V L S G V A L A ccnaaccagcacaantacgtcacggcagcccagctggtgtcgctgagctatcaatccggt cgggcgcgganacgntgtgggtccgcgcggatgacggcaccgtctgggatcatggtcaag RARXXCGSARMTAPSGIMVK tgctttcacngtgactgcgccggtggacacgggtccggtggtgaccccgaccaactcaag C F H X D C A G G H G S G G D P D Q L K cacgctgtccgttcaaggccagacgttctcggtgtcttcgctcttcacctattctgatcc V F A L H L F -H A V R S R P D V L G tcggtgagaggntggagagt, GENINO: 25

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3'5' GCA2a Frame 2

 $\verb|nttcnaaananttacgnccaaagcgcgcaaattaaccntcaataaaaggaacaaaaagtgn|$ F X X X T X K A R K L T X N K R N K K X agctncaccgcggttgccgccngcttaaagnccnaccgacgtggacatatcaatgaaagg S X T A V A A X L K X X R R G H I N E R aatcggcgcaaatgcgggcacggtgtggggggggtggtcgaatgctttcccngtaaatgnt N R R K C G H G V G E W S N A F X V N X $\verb|cccctngacacggtacccgttgagacggtgtnnaacatcgttgcgccccacggcnagagn|\\$ P X D T V P V E T V X N I V A P H G X X tatgcggnttngtcgctgtgcacntacagtgatccgttcgacagtgctgcgacccaatat Y A X X S L C X Y S D P F D S A A T Q Y gacgtttgggacacgggcacaggtgcggccantttgtcttgagcggggtggcacttgcac D V W D T G T G A A X L S - A G W H L H cnaaccagcacaantacgtcacggcagcccagctggtgtcgctgagctatcaatccggtc $\hbox{\tt X} \quad \hbox{\tt T} \quad \hbox{\tt S} \quad \hbox{\tt T} \quad \hbox{\tt X} \quad \hbox{\tt T} \quad \hbox{\tt S} \quad \hbox{\tt R} \quad \hbox{\tt Q} \quad \hbox{\tt P} \quad \hbox{\tt S} \quad \hbox{\tt W} \quad \hbox{\tt C} \quad \hbox{\tt R} \quad - \quad \hbox{\tt A} \quad \hbox{\tt I} \quad \hbox{\tt N} \quad \hbox{\tt P} \quad \hbox{\tt V}$ gggcgcgganacgntgtgggtccgcgcggatgacggcaccgtctgggatcatggtcaagt GRGXXVGPRG-RHRLGSWSS gctttcacngtgactgcgccggtggacacgggtccggtggtgaccccgaccaactcaagc A F X V T A P V D T G P V V T P T N S S ${\tt acgctgtccgttcaaggccagacgttctcggtgtcttcgctcttcacctattctgatcct}$ T L S V Q G Q T F S V S S L F T Y S D P cggtgagaggntggagagt

3'5' GCA2a Frame 3

nttcnaaananttacgnccaaagcgcgcaaattaaccntcaataaaaggaacaaaaagtgna

X K X L X P K R A N - X S I K G T K S X gctncaccgcggttgccgccngcttaaagnccnaccgacgtggacatatcaatgaaagga A X P R L P X A - X X T D V D I S M K G ${\tt atcggcgcaaatgcgggcacggtgtgggggggtggtcgaatgctttcccngtaaatgntc}$ I G A N A G T V W G S G R M L S X - M X $\verb"ccctngacacggtacccgttgagacggtgtnnaacatcgttgcgccccacggcnagagnt"$ P X T R Y P L R R C X T S L R P T X R X ${\tt atgcggnttngtcgctgtgcacntacagtgatccgttcgacagtgctgcgacccaatatg}$ M R X X R C A X T V I R S T V L R P N M T F G T R A Q V R P X C L E R G G T C T naaccagcacaantacgtcacggcagcccagctggtgtcgctgagctatcaatccggtcg X P A Q X R H G S P A G V A E L S I R S ggcgcgganacgntgtgggtccgcgcggatgacggcaccgtctgggatcatggtcaagtg G A X T X W V R A D D G T V W D H G Q V ctttcacngtgactgcgccggtggacacgggtccggtggtgaccccgaccaactcaagca LSX-LRRWTRVRW-PRPTQA cgctgtccgttcaaggccagacgttctcggtgtcttcgctcttcacctattctgatcctc R C P F K A R R S R C L R S S P I L I L ES (SERID NO:28)

GCA3b

GATCCGACCAGCAATCAGGCGGAGCTGCAGCACCTGAAAAACGACCTTCT - 50 CTCGGCACTGCTGGGTATTTCACGCAACCGCTCTGCGCTTGGCGGGAAAC -100ACCGACGCGCTTGAAGGCTTACCGGACGACACGCCGCCAGCCTTGATTCG AATGCATCTGGAGTACTTGCGCAGTCAGGATTCCGAGCAGCGCGCCAAGC -200TGTCCGAACTGGATCAGCAACGGGTGCAGAAGGTCGCGGAGACCAGGACG -250ATCGACGCCAGCATCGCGAAGATTGAAGCTTTGCTGCGGTGCTGCAGGAN -300 CGGGTCGGGGTTCGCAAGTACCTGGCGGACAGGGAGTACGGCTCAAAGCT -350GCAATATTCGCAGGAACTCCAGGAACTGGTCGGGATGCAGCAGGACATCC -400TGGTGCAACGGAGCAAAGCTCGAGGAAACCAATGCGGNTTGTCGCCGCAC -450 -500 TTCGACGAAAACCCGCGGNAAGCTTCGTCTNNGAATAACCGGCACCCGNC TGTTCCNACGATCTTGGCCCAAGGGGACGCAAAAAAGGGCCGGCAAGNCC -550TCAAAGGACCAAGGGNGTTTTAAAANCCGAGCACCCGGGACCCAACCTTT -600 -650AAAAANCNTTGGCGGCCCCCATTCGACGGNGTGGNGGCAACAAATTGGGC CGNGCCCCATTT -662 SEQ ID NO:10

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5'3' GCA3b Frame 1

gatccgaccagcaatcaggcggagctgcagcacctgaaaaacgaccttctctcggcactg D P T S N Q A E L Q H L K N D L L S A L ctgggtatttcacgcaaccgctctgcgcttggcgggaaacaccgacgcgcttgaaggctt L G I S R N R S A L G G K H R R A - R L accggacgacacgccgccagccttgattcgaatgcatctggagtacttgcgcagtcagga T G R H A A S L D S N A S G V L A Q S G ttccgagcagcgcccaagctgtccgaactggatcagcaacgggtgcagaaggtcgcgga F R A A R Q A V R T G S A T G A E G R G gaccaggacgatcgacgccagcatcgcgaagattgaagctttgctgcggtgctgcaggan D Q D D R R Q H R E D - S F A A V L Q X cgggtcggggttcgcaagtacctggcggacagggagtacggctcaaagctgcaatattcg R V G V R K Y L A D R E Y G S K L Q Y S caggaactccaggaactggtcgggatgcagcaggacatcctggtgcaacggagcaaagct Q E L Q E L V G M Q Q D I L V Q R S K A cgaggaaaccaatgcggnttgtcgccgcacttcgacgaaaacccgcggnaagcttcgtct RGNQCXLSPHFDENPRXASS nngaataaccggcacccgnctgttccnacgatcttggcccaaggggacgcaaaaaagggc X N N R H P X V X T I L A Q G D A K K G cggcaagncctcaaaggaccaagggngttttaaaanccgagcacccgggacccaaccttt R Q X L K G P R X F - X P S T R D P H, (SEO ID AD: 38) aaaaancnttggcggcccccattcgacggngtggnggcaacaaattgggccgngccccat ${f T}$ N Х tt, (SEQ I) NO: 291

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5'3' GCA3b Frame 2

I R P A I R R S C S T - K T T F S R H C tgggtatttcacgcaaccgctctgcgcttggcgggaaacaccgacgcgcttgaaggctta WVFHATALRLAGNTDALEGL ccggacgacacgccgccagccttgattcgaatgcatctggagtacttgcgcagtcaggat P D D T P P A L I R M H L E Y L R S Q D tecgageagegegecaagetgteegaactggateageaacgggtgeagaaggtegeggag S E Q R A K L S E L D Q Q R V Q K V A E accaggacgatcgacgccagcatcgcgaagattgaagctttgctgcggtgctgcagganc T R T I D A S I A K I E A L L R C C R X $\tt gggtcggggttcgcaagtacctggcggacagggagtacggctcaaagctgcaatattcgc$ G S G F A S T W R T G S T A Q S C N I R aggaactccaggaactggtcgggatgcagcaggacatcctggtgcaacggagcaaagctc R N S R N W S G C S R T S W C N G A K L gaggaaaccaatgcggnttgtcgccgcacttcgacgaaaacccgcggnaagcttcgtctn E E T N A X C R R T S T K T R X K L R X ngaataaccggcacccgnctgttccnacgatcttggcccaaggggacgcaaaaaagggcc X I T G T X L F X R S W P K G T Q K R A ggcaagncctcaaaggaccaagggngttttaaaanccgagcacccgggacccaaccttta G K X S K D Q G X F K X R A P G T Q P aaaancnttggcggccccattcgacggngtggnggcaacaaattgggccgngccccatt SW X D Z X X X G G P H S T X W X Q Q I G P X P I + (SEQ ID NO: 31)

DOLBUZZZ "D8180

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5'3' GCA3b Frame 3

gatccgaccagcaatcaggcggagctgcagcacctgaaaaacgaccttctctctggcactgct

S D Q Q S G G A A A P E K R P S L G T A $\tt gggtatttcacgcaaccgctctgcgcttggcgggaaacaccgacgcgcttgaaggcttac$ G Y F T Q P L C A W R E T P T R L K A Y cggacgacacgccgccagccttgattcgaatgcatctggagtacttgcgcagtcaggattR T T R R Q P - F E C I W S T C A V R I ccgagcagcgcccaagctgtccgaactggatcagcaacgggtgcagaaggtcgcggaga PSSAPSCPNWISNGCRRSRR ccaggacgatcgacgccagcatcgcgaagattgaagctttgctgcggtgctgcaggancg P G R S T P A S R R L K L C C G A A G X ggtcggggttcgcaagtacctggcggacagggagtacggctcaaagctgcaatattcgca GRGSQVPGGQGVRLKAAIFA ggaactccaggaactggtcgggatgcagcaggacatcctggtgcaacggagcaaagctcg G T P G T G R D A A G H P G A T E Q S S aggaaaccaatgcggnttgtcgccgcacttcgacgaaaacccgcggnaagcttcgtctnnR K P M R X V A A L R R K P A X S F V X gaataaccggcacccgnctgttccnacgatcttggcccaaggggacgcaaaaaagggccg E - P A P X C S X D L G P R G R K K G P gcaagncctcaaaggaccaagggngttttaaaanccgagcacccgggacccaacctttaa

A X P Q R T K X V L K X E H P G P N L - SEL I NO.

aaancnttggcggccccattcgacggngtggnggcaacaaattgggccgngccccattt G

K X L A A P I R R X X G N K L

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3'5' GCA3b Frame 1

aaatggggcncggcccaatttgttgccnccacnccgtcgaatgggggccgccaangnttt ttaaaggttgggtcccgggtgctcggnttttaaaacncccttggtcctttgaggncttgc L K V G S R V L X F - N X L G P L R X C cggcccttttttgcgtccccttgggccaagatcgtnggaacagncgggtgccggttattc R P F F A S P W A K I X G T X G C R L F ${\tt nnagacgaagcttnccgcgggttttcgtcgaagtgcggcgacaanccgcattggtttcct}$ X D E A X R G F S S K C G D X P H W F P cgagctttgctccgttgcaccaggatgtcctgctgcatcccgaccagttcctggagttcc R A L L R C T R M S C C I P T S S W S S tgcgaatattgcagctttgagccgtactccctgtccgccaggtacttgcgaaccccgacc CEYCSFEPYSLSARYLRTPT cgntcctgcagcaccgcagcaaagcttcaatcttcgcgatgctggcgtcgatcgtcctgg X S C S T A A K L Q S S R C W R R S S W tetecgegacettetgcaccegttgctgatecagtteggacagettggegegetgetegg S P R P S A P V A D P V R T A W R A A R aatcctgactgcgcaagtactccagatgcattcgaatcaaggctggcggcgtgtcgtccg N P D C A S T P D A F E S R L A A C R P gtaagccttcaagcgcgtcggtgtttcccgccaagcgcagagcggttgcgtgaaataccc V S L Q A R R C F P P S A E R L R E agcagtgccgagagaaggtcgtttttcaggtgctgcagctccgcctgattgctggtcgga AERRSFFRCCS

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3'5' GCA3b Frame 2

aaatggggcncggcccaatttgttgccnccacnccgtcgaatgggggccgccaangntttt N G X R P N L L X P X R R M G A A X X F taaaggttgggtcccgggtgctcggnttttaaaacncccttggtcctttgaggncttgcc - R L G P G C S X F K X P L V L - X L A ggcccttttttgcgtccccttgggccaagatcgtnggaacagncgggtgccggttattcn G P F L R P L G P R S X E Q X G A G Y X ${\tt nagacgaagcttnccgcgggttttcgtcgaagtgcggcgacaanccgcattggtttcctc}$ X T K L X A G F R R S A A T X R I G F L gagetttgetcegttgeaccaggatgtcctgctgcatcccgaccagttcctggagttcct E L C S V A P G C P A A S R P V P G V P gcgaatattgcagctttgagccgtactccctgtccgccaggtacttgcgaaccccgaccc ANIAALSRTPCPPGTCEPRP gntcctgcagcaccgcagcaaagcttcaatcttcgcgatgctggcgtcgatcgtcctggt X P A A P Q Q S F N L R D A G V D R P G ctccgcgaccttctgcacccgttgctgatccagttcggacagcttggcgcgctgctcgga L R D L L H P L L I Q F G Q L G A L L G ${\tt atcctgactgcgcaagtactccagatgcattcgaatcaaggctggcggcgtgtcgtccgg}$ I L T A Q V L Q M H S N Q G W R R V V R taagccttcaagcgcgtcggtgtttcccgccaagcgcagagcggttgcgtgaaataccca - A F K R V G V S R Q A Q S G C V K Y P 1 (SEQ ID NO: 57) gcagtgccgagagaaggtcgtttttcaggtgctgcagctccgcctgattgctggtcggat A V P R E G R F S G A A A P P

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3'5' GCA3b Frame 3

aaatggggcncggcccaatttgttgccnccacnccgtcgaatgggggccgccaangnttttt M G X G P I C C X H X V E W G P P X X F ${\tt aaaggttgggtcccgggtgctcggnttttaaaacncccttggtcctttgaggncttgccg}$ K G W V P G A R X L K X P W S F E X L P gcccttttttgcgtccccttgggccaagatcgtnggaacagncgggtgccggttattcnn A L F C V P L G Q D R X N X R V P V I X ${\tt agacgaagcttnccgcgggttttcgtcgaagtgcggcgacaanccgcattggtttcctcg}$ R R S X P R V F V E V R R Q X A L V S S agetttgeteegttgeaccaggatgteetgetgeatecegaccagtteetggagtteetg S F A P L H Q D V L L H P D Q F L E F L cgaatattgcagctttgagccgtactccctgtccgccaggtacttgcgaaccccgacccg R I L Q L - A V L P V R Q V L A N P D P ${\tt ntcctgcagcaccgcagcaaagcttcaatcttcgcgatgctggcgtcgatcgtcctggtc}$ X L Q H R S K A S I F A M L A S I V L V tecgcgacettetgcacecgttgctgatecagtteggacagettggegegetgeteggaa S A T F C T R C - S S S D S L A R C S E tectgactgegeaagtactecagatgeattegaateaaggetggeggegtgtegteeggt S - L R K Y S R C I R I K A G G V S S G aagccttcaagcgcgtcggtgtttcccgccaagcgcagagcggttgcgtgaaatacccag K P S S A S V F P A K R R A V A 1 (SEDIANO:3 cagtgccgagagaaggtcgtttttcaggtgctgcagctccgcctgattgctggtcggatc

GCA4

ACTCTCNNGCCTCTCACCGAAGATAGCCGGCAAGGACTGGCGNGAACANN -50 -100

QLRL

GCGCGCTGGACTATCNCTAAAGGGTCTCCNACNACGTCCANCCGGACNAG CTGACCTCGTTTCCNCNAAGCGTGAAACTGAAGGCCGGTGAAACCNTCNT -150-200

GTTCGCCTNGATCACCTACTAGTCGCGCGCCNNGCGCGACAGGATCAACG CCAAGGTGATGGCCGATCCCCGCCTGGCGTCGTCGATGGATC -242

CREKVVFQVL

SEO ID NO:11

5'3' GCA4 Frame 1

cgcgctggactatenetaaagggteteenacnacgteeaneeggacnagetgacetegtt
R A G L X L K G L X X R P X G X A D L V

teenenaagegtgaaactgaaggeeggtgaaacententgttegeetngateacetacta
s X K R E T E G R - N X X V R X D H L L

gtegegegeenngegegacaggateaacgccaaggtgatggeegateeeegeetggegte
V A R X A R Q D Q R Q G D G R S P P G V

gtegatggate
V D G CHO FO NO!40

5'3' **GCA4** Frame 2

cgcgctggactatenctaaagggteteenacnacgteeanceggacnagetgacetegtte

A L D Y X - R V S X X V X P D X L T S F

cenenaagegtgaaactgaaggeeggtgaaacententgttegeetngateacetactag

X X S V K L K A G E T X X F A X I T Y
tegegegeenngegegacaggateaacgccaaggtgatggeegateceegcetggegteg

S R A X R D R I N A K V M A D P R L A S

tegatggate

S M D G D J J NO! 42

5'3' GCA4 Frame 3

cgcgctggactatenctaaagggtctccnacnacgtccanccggacnagctgacctcgtttc

R W T I X K G S X X T S X R X S - P R F

cncnaagcgtgaaactgaaggccggtgaaaccntcntgttcgcctngatcacctactagt

X X A - N - R P V K X X C S P X S P T S

cgcgcgccnngcgcgacaggatcaacgccaaggtgatggccgatccccgcctggcgtcgt

R A X X A T G S T P R - W P I P A W R R

cgatggatc, GROID NO: 43

R W I GROID NO: 44

P 30

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3'5' GCA4 Frame 1

3'5' GCA4 Frame 2

gatccatcgacgacgccaggcggggatcggccatcaccttggcgttgatcctgtcgcgcnn

I H R R R Q A G I G H H L G V D P V A X

ggcgcgcgactagtaggtgatcnaggcgaacanganggtttcaccggccttcagtttcac
G A R L V G D X G E X X G F T G L Q F H

gcttngnggaaacgaggtcagctngtccggntggacgtngtnggagaccctttagngata
A X X K R G Q X V R X D X X G D P L X I

gtccagcgcg
V Q R

3'5' **GCA4** Frame 3

gatccatcgacgacgccaggcggggatcggccatcaccttggcgttgatcctgtcgcgcnng

S I D D A R R G S A I T L A L I L S R X

gcgcgcgactagtaggtgatcnaggcgaacanganggtttcaccggccttcagtttcacg

A R D - - V I X A N X X V S P A F S F T

cttngnggaaacgaggtcagctngtccggntggacgtngtnggagaccctttagngatag

L X G N E V S X S X W T X X E T L - X
tccagcgcg

S S A Choll No! 48

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GATCCGCTCGATGCCCAGGCCCAGTACAGCGAACTGTTCGCCCATGGCCG -50 ${\tt CGCCACGTCACTGTTGCTATTCGAACATGTTCACGGTGAATCCCGTGACC}$ -100 -150 GCGGCCAGGCGATGGTGGACCTGCTGGCGCAGTACGAGCAGCACGGTTTG CAGTTAAACAGCCGCGAATTACCGGACCACCTGCCGCTGTATCTGGAGTA -200CCTGTCGCAGCTGCCGCAAGGCGAAGCCGTGGAAGGTTTGAAAGATATCG -250CGCCGATTCTGGCATTGCTGAGCGCGCGTCTGCAACAGCGTGAAAGCCGT -350TATGCCGTGATGTTTGATCTGCTGCTGAAATTGGCCGATACCGCTATCGA CAGCGACAAAGTGGCGGAAAAAATTGCCGACGAAGCGCGCGATGATACGC -400 CGCAGGCGCTGGATGCTGTTTGGGAAGAAGAGCAGGTTAAATTCTTTGCT -450-500GACAAAGGCTGCGGCGATTCAGCAATCACTGCTCATCAGCGTCGCTTTGC -550 CGGTGCCGTCGCCGCAATATCTGAATATCCTCGGTGAGAGGCTGGAGA SEQ ID NO:12 -552

5'3' **GCA5** Frame 1

gatccgctcgatgcccaggcccagtacagcgaactgttcgcccatggccgcgccacgtca D P L D A Q A Q Y S E L F A H G R A T S ctgttgctattcgaacatgttcacggtgaatcccgtgaccgcggccaggcgatggtggac L L L F E H V H G E S R D R G Q A M V D ctgctggcgcagtacgagcagcacggtttgcagttaaacagccgcgaattaccggaccac LLAQYEQHGLQLNSRELPDH ctgccgctgtatctggagtacctgtcgcagctgccgcaaggcgaagccgtggaaggtttg L P L Y L E Y L S Q L P Q G E A V E G L ${\tt aaagatatcgcgccgattctggcattgctgagcgcgcgtctgcaacagcgtgaaagccgt}$ K D I A P I L A L L S A R L Q Q R E S R tatgccgtgatgtttgatctgctgctgaaattggccgataccgctatcgacagcgacaaa Y A V M F D L L L K L A D T A I D S D K gtggcggaaaaaattgccgacgaagcgcgcgatgatacgccgcaggcgctggatgctgtt V A E K I A D E A R D D T P Q A L D A V tgggaagaagagcaggttaaattctttgctgacaaaggctgcggcgattcagcaatcact WEEEQVKFFADKGCGDSAIT gctcatcagcgtcgctttgccggtgccgtcgcgcaatatctgaatatcctcggtgag VAPQYLNILGE R F A G A R aggctggagagt, BEQ ID 119:49

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5'3' GCA5 Frame 2

gatccgctcgatgcccaggcccagtacagcgaactgttcgcccatggccgcgccacgtcac I R S M P R P S T A N C S P M A A P R H tgttgctattcgaacatgttcacggtgaatcccgtgaccgcggccaggcgatggtggacc C C Y S N M F T V N P V T A A R R W W T tgctggcgcagtacgagcacggtttgcagttaaacagccgcgaattaccggaccacc C W R S T S S T V C S - T A A N Y R T T tgccgctgtatctggagtacctgtcgcagctgccgcaaggcgaagccgtggaaggtttga CRCIWSTCRSCRKA·KPWKV- ${\tt aagatatcgcgccgattctggcattgctgagcgcgccgtctgcaacagcgtgaaagccgtt}$ KISRRFWHC-ARVCNSVKAV atgccgtgatgtttgatctgctgctgaaattggccgataccgctatcgacagcgacaaag MP-CLICC-NWPIPLSTATK tggcggaaaaaattgccgacgaagcgcgcgatgatacgccgcaggcgctggatgctgttt W R K K L P T K R A M I R R R W M L F gggaagaagagcaggttaaattctttgctgacaaaggctgcggcgattcagcaatcactg G K K S R L N S L L T K A A A I Q Q S L ctcatcagcgtcgctttgccggtgccgtcgcgcaatatctgaatatcctcggtgaga ggctggagagt SEQ IN 10:51 R, COROLINO: 52)

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5'3' GCA5 Frame 3

 Gate construction of the strength of the st

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3'5' GCA5 Frame 1

actctccagcctctcaccgaggatattcagatattgcggcgcgacggcaccggcaaagcg TLQPLTEDIQILRRDGTGKA acgctgatgagcagtgattgctgaatcgccgcagcctttgtcagcaaagaatttaacctg T L M S S D C - I A A A F V S K E F N L ctcttcttcccaaacagcatccagcgcctgcggcgtatcatcgcgcgcttcgtcggcaat L F F P N S I Q R L R R I I A R F V G N $\verb|ttttccgccactttgtcgctgtcgatagcggtatcggccaatttcagcagcagatcaaa|$ F F R H F V A V D S G I G Q F Q Q I K catcacggcataacggctttcacgctgttgcagacgcgcgctcagcaatgccagaatcgg H H G I T A F T L L Q T R A Q Q C Q N R cgcgatatctttcaaaccttccacggcttcgccttgcggcagctgcgacaggtactccag R D I F Q T F H G F A L R Q L R Q V L Q atacagcggcaggtggtccggtaattcgcggctgtttaactgcaaaccgtgctgctcgta I Q R Q V V R - F A A V - L Q T V L L V ctgcgccagcaggtccaccatcgcctggccgcggtcacgggattcaccgtgaacatgttc L R Q Q V H H R L A A V T G F T V N M F gaatagcaacagtgacgtggcgcggccatgggcgaacagttcgctgtactgggcctgggc QQ-RGAAMGEQFAVLGLG atcgagcggatc (SEOIL 10:55)

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3'5' GCA5 Frame 2

actctccagcctctcaccgaggatattcagatattgcggcgcgacggcaccggcaaagcga L S S L S P R I F R Y C G A T A P A K R cgctgatgagcagtgattgctgaatcgccgcagcctttgtcagcaaagaatttaacctgc R - - A V I A E S P Q P L S A K N L T C tettetteccaaacagcatecagegectgeggegtateategegegettegteggeaatt S S S Q T A S S A C G V S S R A S S A I ttttccgccactttgtcgctgtcgatagcggtatcggccaatttcagcagcagatcaaac F S A T L S L S I A V S A N F S S R S N atcacggcataacggctttcacgctgttgcagacgcgctcagcaatgccagaatcggc ITA-RLSRCCRRALSNARIG gcgatatctttcaaaccttccacggcttcgccttgcggcagctgcgacaggtactccaga A I S F K P S T A S P C G S C D R Y S R tacagcggcaggtggtccggtaattcgcggctgtttaactgcaaaccgtgctgctcgtac Y S G R W S G N S R L F N C K P C C S Y tgcgccagcaggtccaccatcgcctggccgcggtcacgggattcaccgtgaacatgttcg C A S R S T I A W P R S R D S P - T C S aatagcaacagtgacgtggcgcgccatgggcgaacagttcgctgtactgggcctgggca NSNSDVARPWANSSLYWAWA s s G GRALL NO:57

3'5' GCA5 Frame 3

actetecageeteteacegaggatatteagatattgeggegegaeggeaceggeaaagegae S P A S H R G Y S D I A A R R H R Q S D gctgatgagcagtgattgctgaatcgccgcagcctttgtcagcaaagaatttaacctgct ADEQ-LLNRRSLCQQRI-PA $\verb"cttcttcccaaacagcatccagcgccttgcggcgtatcatcgcgcgcttcgtcggcaattt$ L L P K Q H P A P A A Y H R A L R R Q F tttccgccactttgtcgctgtcgatagcggtatcggccaatttcagcagcagatcaaaca F P P L C R C R - R Y R P I S A A D Q T teacggcataacggctttcacgctgttgcagacgcgcgctcagcaatgccagaatcggcg S R H N G F H A V A D A R S A M P E S A cgatatctttcaaaccttccacggcttcgccttgcggcagctgcgacaggtactccagat R Y L S N L P R L R L A A A A T G T P D ${\tt acagcggcaggtggtccggtaattcgcggctgtttaactgcaaaccgtgctgctcgtact}$ T A A G G P V I R G C L T A N R A A R T gcgccagcaggtccaccatcgcctggccgcggtcacgggattcaccgtgaacatgttcga A P 'A G P P S P G R G H G I H R E H V R atagcaacagtgacgtggcgcggccatgggcgaacagttcgctgtactgggcctgggcat I A T V T W R G H G R T V R C T G P G H cgagcggatc R A DAGERTA NO: 58)

GCA7

GATCCTNACACANTAGCCCGTGGACGCATTTGCGTCGACCCTCATANGGA -50
AGCGATACGAGGCGGGTNAAAGTGAACATCCGCCGAGCACGCAGCGACG
CCTCCGCTCACCGTCNGCGCAGTACTTCCTCGGGTCGCCGCGCCTAGCAC -150
TCTGCGCCGTGACATCAANCCGTGAACCCACGGGAGACTTTGCGCCGCNA -200
AGGGATGAGTCCACTATTAGATGACGCATGGCTACGAGCCNATCCTCGGT
GANAAGCTGGAGAGT -265 SEQ ID NO:13

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5'3' GCA7 Frame 1

5'3' GCA7 Frame 2

gatectnacacantagecegtggaegeatttgegtegaeceteatanggaagegataegag

I X T X - P V D A F A S T L I X K R Y E

gegggtnaaagtgaacateegeegageacgeagegaegeeteegeteacegtengegea

A G X S E H P P S T A A T P P L T V X A

gtactteetegggtegeegeetageaetetgegeegtgaeateaancegtgaaceeae

V L P R V A A P S T L R R D I X P - T H

gggagaetttgegeegenaagggatgagteeaetattagatgaegeatggetaegageen

G R L C A X K G - V H Y - M T H G Y E X

atceteggtganaagetggagagt GALL NO: 621

I L G X K L E S GALL NO: 621

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5'3' **GCA7** Frame 3

gatectnacacantagecegtggaegeatttgegtegaeeeteatanggaagegataegagg
S X H X S P W T H L R R P S X G S D T R

cgggtnaaagtgaacateegeegageaeggeaeggeeteegeteaeegtengegeag
R X K V N I R R A R Q R R L R S P X A Q

tactteetegggtegeegegeetageaetetgegeegtgaeateaaneegtgaaeeeaeg
Y F L G S P R L A L C A V T S X R E P T

ggagaetttgegeegenaagggatgagteeaetattagatgaegeatggetaegageena
G D F A P X R D E S T I R - R M A T S X

teeteggtganaagetggagagt
S S V X S W R

3'5' GCA7 Frame 1

actctccagcttntcaccgaggatnggctcgtagccatgcgtcatctaatagtggactca

T L Q L X T E D X L V A M R H L I V D S

tcccttngcggcgcaaagtctcccgtgggttcacggnttgatgtcacggcgcagagtgct

S L X G A K S P V G S R X D V T A Q S A

aggcgcggcgacccgaggaagtactgcgcngacggtgagcggaggcgtcgctgccgtgct

R R G D P R K Y C X D G E R R R R C R A

cggcggatgttcactttnacccgcctcgtatcgcttccntatgagggtcgacgcaaatgc

R R M F T X T R L V S L X Y E G R R K C

gtccacgggctantgtgtnaggatc SWAA W: 66

V H G L X C X D GWAA W R H L I V D S L X Y E G R R K C

3'5' **GCA7** Frame 2

actetecagettnteacegaggatnggetegtagecatgegteatetaatagtggacteat

L S S X S P R X G S - P C V I - - W T H

ccettngeggegeaaagteteeegtgggtteacggnttgatgteacggegagagtgeta

P X A A Q S L P W V H X L M S R R R V L

ggegggggaceegaggaagtaetgegengaeggtgageggaggegtegetgeegtgete

G A A T R G S T A X T V S G G V A A V L

ggeggatgtteaetttnaceegeetegtategetteentatgagggtegaegeaaatgeg

G G C S L X P A S Y R F X M R V D A N A

tecaegggetantgtgtnaggate

S T G X C X R I

3'5' **GCA7** Frame 3

GCA10

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GATCCGGCCNCGCACGANCTTACCGGTNAAAACTTCCNCNCCNATAATAT -50
TTGCCGCGCGAGCCGCCCTGANGCTCTCGGCGTAACTCCGGATGCACGGG -100
GGACCGTGACGGTTGTANTGCCCTGGCTTTTCTCAGCNGAAATCTGCACA -150
GCCATCTTCCGATCGATCTGGCGCAGGTGGGGCCNCAAAACGGTGGGC -200
ATCTCCAAACCGCAGGAACGTGTTTTGCAGGATGTCGAACATCATCCACG -250
CTTCGGTNCCCAACGGCTACTTCGCCCGGTACCGGGCCATGTCATCCTCG -300
GTGANAAGCTGGANANT -317 SEQ ID NO:14

5'3' GCA10 Frame 1 gatccggccncgcacgancttaccggtnaaaacttccncnccnataatatttgccgcgcg D P A X H X L T G X N F X X X N I C R A ${\tt agccgccctgangctctcggcgtaactccggatgcacgggggaccgtgacggttgtantg}$ S R P X A L G V T P D A R G T V T V V X ccctggcttttctcagcngaaatctgcacagccatcttccgatcgatctggcgcaggtgg PWLFSXEICTAIFRSIWRRW ggcggcncaaaacggtgggcatctccaaaccgcaggaacgtgttttgcaggatgtcgaac G G X K R W A S P N R R N V F C R M S N atcatccacgcttcggtncccaacggctacttcgcccggtaccgggccatgtcatcctcg I I H A S X P N G Y F A R Y R A M S S S gtganaagctgganant, (Short) No: 69 x (500 Is No: 70) V X S W X

5'3' GCA10 Frame 2

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gatccggccncgcacgancttaccggtnaaaacttccncnccnataatatttgccgcgcga I R X R T X L P X K T S X X I I F A A R gccgccctgangctctcggcgtaactccggatgcacgggggaccgtgacggttgtantgc AALXLSA-LRMHGGP-RLXC cctggcttttctcagcngaaatctgcacagccatcttccgatcgatctggcgcaggtggg P G F S Q X K S A Q P S S D R S G A G G gcggcncaaaacggtgggcatctccaaaccgcaggaacgtgttttgcaggatgtcgaaca A X Q N G G H L Q T A G T C F A G C R T ${\tt tcatccacgcttcggtncccaacggctacttcgcccggtaccgggccatgtcatcctcgg}$ S S T L R X P T A T S P G T G P C H P R tganaagctgganant, CELIA NO:71

5'3' GCA10 Frame 3 gatccggccncgcacgancttaccggtnaaaacttccncnccnataatatttgccgcgcgag S G X A R X Y R X K L X X X - Y L P R E $\verb|ccgccctgangctctcggcgtaactccggatgcacgggggaccgtgacggttgtantgcc|$ PP-XSRRNSGCTGDRDGCXA ctggcttttctcagcngaaatctgcacagccatcttccgatcgatctggcgcaggtgggg LAFLSXNLHSHLPIDLAQVG cggcncaaaacggtgggcatctccaaaccgcaggaacgtgttttgcaggatgtcgaacat $\stackrel{-}{\mathsf{R}}$ X K T V G I S K P Q E R V L Q D V E H catccacgcttcggtncccaacggctacttcgcccggtaccgggccatgtcatcctcggt HPRFGXQRLLRPVPGHVILG ganaagctgganant (SEQ TI NO173) X K L

3'5' GCA10 Frame 1 ${\tt antntccagcttntcaccgaggatgacatggcccggtaccgggcgaagtagccgttgggn}$ X X Q L X T E D D M A R Y R A K - P L X accgaagcgtggatgatgttcgacatcctgcaaaacacgttcctgcggtttggagatgcc TEAWMMFDILQNTFLRFGDA caccgttttgngccgcccacctgcgccagatcgatcggaagatggctgtgcagatttcn H R F X P P H L R Q I D R K M A V Q I X gctgagaaaagccagggcantacaaccgtcacggtcccccgtgcatccggagttacgccg A E K S Q G X T T V T V P R A S G V T P agagcntcagggcggctcgcggcaaatattatnggngnggaagttttnaccggtaagn R X S G R L A R Q I L X X X K F X P V X tcgtgcgnggccggatc (glots No:75) G 1980 LD NO: 76)

3'5' **GCA10** Frame 2

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antntccagcttntcaccgaggatgacatggcccggtaccgggcgaagtagccgttgggna X S S X S P R M T W P G T G R S S R W X ${\tt ccgaagcgtggatgatgttcgacatcctgcaaaacacgttcctgcggtttggagatgccc}$ PKRG-CSTSCKTRSCGLEMP accgttttgngccgccccacctgcgccagatcgatcggaagatggctgtgcagatttcng T V L X R P T C A R S I G R W L C R F X ctgagaaaagccagggcantacaaccgtcacggtcccccgtgcatccggagttacgccga L R K A R A X Q P S R S P V H P E L R R gagcntcagggcggctcgcgggcaaatattatnggngnggaagttttnaccggtaagnt E X Q G G S R G K Y Y X X G S F X R - X cgtgcgnggccggatc RAXPD/ (SED IN NO! 77)

3'5' GCA10 Frame 3

antntccagcttntcaccgaggatgacatggcccggtaccgggcgaagtagccgttgggnac X P A X H R G - H G P V P G E V A V G X cgaagcgtggatgatgttcgacatcctgcaaaacacgttcctgcggtttggagatgccca ccgttttgngccgccccacctgcgccagatcgatcggaagatggctgtgcagatttcngc P F X A A P P A P D R S E D G C A D F X tgagaaaagccagggcantacaaccgtcacggtcccccgtgcatccggagttacgccgag - E K P G X Y N R H G P P C I R S Y A E ${\tt agentcagggcggctcgcgcgcaaatattatnggngnggaagttttnaccggtaagntc}$ S X R A A R A A N I X X X E V X T G K X gtgcgnggccggatc 1/ (SEQ ID NO: 18) V X G R

GCA12

ACTCTCCAGCCTCGCACCGAGGATCAGGGCGTCGTCGACCTCGACCT -50GACCGCCTCCCCNCCGCTGCTCTCGATCGGCGGCCAGACCTACACCANCG -100ACGTAGATCAAGCGCGTGGTGCGCGCGCCNACNAGCANCANCTAANTCAA -150GGCCTCGCTGCATCCCGCCAATCCAGCGCTCAGCTTCGCGGGAATTGCGC -200 GANCGCTTTTGCGCGTCNCGAGTNACCGCATACACACCTGCCGTCCCTGC -250GAAAGCAAGGACCCATACTCCGCNGCGGGTGTTGTTGACGGGACTCGTCA -300TGGCGGCAACGCACAACGTNNAACTTCTGTGGTTATGGATC -341 SEQ ID NO:15

5'3' GCA12 Frame 1

5'3' GCA12 Frame 2

L S S L A P R I R A S S T P S T - P P P CONCEGERATE CONCERNATION CONCERNS TO THE PROPERTY OF T

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5'3' GCA12 Frame 3

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actctccagcctcgcaccgaggatcagggcgtcgtcgactccgtcgacctgaccgcctcccc S P A S H R G S G R R R L R R P D R L P ${\tt nccgctgctctcgatcggccagacctacaccancgacgtagatcaagcgcgtggtgc}$ X A A L D R R P D L H X R R R S S A W C geggegenacnageancanetaanteaaggeetegetgeateeegeeaateeagegetea A A X X A X X X S R P R C I P P I Q R S gcttcgcgggaattgcgcgancgcttttgcgcgtcncgagtnaccgcatacacacctgcc A S R E L R X R F C A X R X T A Y T P A gtccctgcgaaagcaaggacccatactccgcngcgggtgttgttgacgggactcgtcatg V P A K A R T H T P X R V L L T G L geggeaacgeacaacgtnnaacttetgtggttatggate, SEQ IJ NO!83) AATHNXXLLWLWINGSEOLINO:94)

3'5' GCA12 Frame 1

gatccataaccacagaagttnnacgttgtgcgttgccgccatgacgagtcccgtcaacaa DP-PQKXXVVRCRHDESRQQ caccegengeggagtatgggteettgetttegeagggaeggeaggtgtgtatgeggtnae H P X R S M G P C F R R D G R C V C G X ${\tt tcgngacgcgcaaaagcgntcgcgcaattcccgcgaagctgagcgctggattggcgggat}$ S X R A K A X A Q F P R S - A L D W R D gcagcgaggccttganttagntgntgctngtngcgccgcgcaccacgcgcttgatctacg A A R P - X X X C X X R R A P R A - S T tcgntggtgtaggtctggccgccgatcgagagcagcggngggggaggcggtcaggtcgacg S X V - V W P P I E S S X G E A V R gagtcgacgacgccctgatcctcggtgcgaggctggagagt (SEQ II) NO:85) E S T T P - S S V R G

3'5' GCA12 Frame 2

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a C

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3'5' GCA12 Frame 3

gatccataaccacagaagttnnacgttgtgcgttgccgccatgacgagtcccgtcaacaaca

GCA19

GATCCGCGCATCCTCTCTGTGGCTCTCGCGGGGTCAGAGGTGGATAAGGC -50
CGGCCGCAAGCTCGGACTTCCCGTCNCAATCNAAGGCTTCTGCGATCNCC -100
ANTACAACTACNACGGCAATCTNACATCACGCAAGATCGCANGCTCNGTC -150
ATCAAGGACGCNGCGGTCNCCNCCCGGCAGGTGCTCNATATNGTGTTGAA -200
NAACACCATCGCTCCTGCAACGGCAAGAAGATCACATGCAAGGTCCACTC -250
GCTGTG -256 SEQ ID NO:16

5'3' GCA19 Frame 1

5'3' GCA19 Frame 2

5'3' GCA19 Frame 3

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B 10

3'5' GCA19 Frame 1

cacagcgagtggaccttgcatgtgatcttcttgccgttgcaggagcgatggtgttnttca H S E W T L H V I F L P L Q E R W C X S a cac natatng agcacct geoggnggngaccg cngcgtcctt gat gac ngagcnt geogagnggngaccg cngcgtcctt gat gac ngagcnt gaT X Y X A P A G X X P X R P - - X S X R ${\tt tcttgcgtgatgtnagattgccgtngtagttgtantggngatcgcagaagccttngattg}$ S C V M X D C R X S C X X D R R S L X L ${\tt ngacgggaagtccgagcttgcggccggccttatccacctctgaccccgcgagagccacag}$ X R E V R A C G R P Y P P L T P R E P Q agaggatgcgcggatc, (SED LINO:95)

R G C A D (SED LINO:96)

3'5' GCA19 Frame 2 ${\tt cacagcgagtggaccttgcatgtgatcttcttgccgttgcaggagcgatggtgttnttcaa}$ T A S G P C M - S S C R C R S D G V X Q ${\tt cacnatatngagcacctgccgggnggngaccgcngcgtccttgatgacngagcntgcgat}$ H X X E H L P X X D R X V L D D X X C D $\verb"cttgcgtgatgtnagattgccgtngtagttgtantggngatcgcagaagccttngattgn"$ L A - C X I A X V V V X X I A E A X D X gacgggaagtccgagcttgcggccggccttatccacctctgaccccgcgagagccacaga D G K S E L A A G L I H L - P R E S H R AR IN CORRESPONDING gaggatgcgcggatc

3'5' GCA19 Frame 3

 ${\tt cacagcgagtggaccttgcatgtgatcttcttgccgttgcaggagcgatggtgttnttcaac}$ Q R V D L A C D L L A V A G A M V X F N ${\tt acnatatngagcacctgccgggnggngaccgcngcgtccttgatgacngagcntgcgatc}$ X I X S T C R X X T X A S L M X E X A I $\verb|ttgcgtgatgtnagattgccgtngtagttgtantggngatcgcagaagccttngattgng|$ L R D X R L P X - L X W X S Q K P X I X acgggaagtccgagcttgcggccggccttatccacctctgaccccgcgagagccacagag G S P S L R P A L S T S D P A R A T E R M R G (SEN IL W: 98)

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Polypeptides and Peptides Associated with GCA

The invention provides polypeptides (e.g., SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:8) associated with GCA, and subsequences of thereof, e.g., peptides. This invention provides immunogenic peptides capable of generating an immune response, particularly antibodies, specifically directed to diagnose or treat GCA. These polypeptides and peptides can also be used to identify the presence of human antibodies that specifically bind to them for the diagnosis of GCA. Polypeptides and peptides of the invention can also be used to generate antibodies that can be used to diagnose or treat GCA.

Polypeptides and peptides of the invention can be isolated from natural sources (e.g., vasculitis lesions), be synthetic, or be recombinantly generated polypeptides. Peptides and proteins can be recombinantly expressed in vitro or in vivo. The peptides and polypeptides of the invention can be made and isolated using any method known in the art, and the invention provides a few exemplary means for generating such proteins.

Polypeptide and peptides of the invention can also be synthesized, whole or in part, using chemical methods well known in the art. See e.g., Caruthers (1980) Nucleic Acids Res. Symp. Ser. 215-223; Horn (1980) Nucleic Acids Res. Symp. Ser. 225-232; Banga, A.K., Therapeutic Peptides and Proteins, Formulation, Processing and Delivery Systems (1995) Technomic Publishing Co., Lancaster, PA. For example, peptide synthesis can be performed using various solid-phase techniques (see e.g., Roberge (1995) Science 269:202; Merrifield (1997) Methods Enzymol. 289:3-13) and automated synthesis may be achieved, e.g., using the ABI 431A Peptide Synthesizer (Perkin Elmer) in accordance with the instructions provided by the manufacturer.

The skilled artisan will recognize that individual synthetic residues and polypeptides incorporating mimetics can be synthesized using a variety of procedures and methodologies, which are well described in the scientific and patent literature, e.g., Organic Syntheses Collective Volumes, Gilman, et al. (Eds) John Wiley & Sons, Inc., NY. Polypeptides incorporating mimetics can also be made using solid phase synthetic procedures, as described, e.g., by Di Marchi, et al., U.S. Pat. No. 5,422,426. Peptides and peptide mimetics of the invention can also be synthesized using combinatorial methodologies. Various techniques for generation of peptide and peptidomimetic libraries are well known, and include, e.g., multipin, tea bag, and split-couple-mix techniques; see, e.g., al-Obeidi (1998) Mol. Biotechnol. 9:205-223; Hruby (1997) Curr. Opin. Chem. Biol. 1:114-119; Ostergaard (1997) Mol. Divers. 3:17-27; Ostresh

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(1996) Methods Enzymol. 267:220-234. Modified peptides of the invention can be further produced by chemical modification methods, see, e.g., Belousov (1997) Nucleic Acids Res. 25:3440-3444; Frenkel (1995) Free Radic. Biol. Med. 19:373-380; Blommers (1994) Biochemistry 33:7886-7896.

Peptides and polypeptides of the invention can also be synthesized and expressed as fusion proteins with one or more additional domains linked thereto for, e.g., producing a more immunogenic peptide, to more readily isolate a recombinantly synthesized peptide, to identify and isolate antibodies and antibody-expressing B cells, and the like. Detection and purification facilitating domains include, e.g., metal chelating peptides such as polyhistidine tracts and histidine-tryptophan modules that allow purification on immobilized metals, protein A domains that allow purification on immobilized immunoglobulin, and the domain utilized in the FLAGS extension/affinity purification system (Immunex Corp, Seattle WA). The inclusion of a cleavable linker sequences such as Factor Xa or enterokinase (Invitrogen, San Diego CA) between the purification domain and GCA-associated peptide or polypeptide can be useful to facilitate purification. For example, an expression vector can include an epitope-encoding nucleic acid sequence linked to six histidine residues followed by a thioredoxin and an enterokinase cleavage site (see e.g., Williams (1995) Biochemistry 34:1787-1797; Dobeli (1998) Protein Expr. Purif. 12:404-14). The histidine residues facilitate detection and purification while the enterokinase cleavage site provides a means for purifying the epitope from the remainder of the fusion protein. Technology pertaining to vectors encoding fusion proteins and application of fusion proteins are well described in the scientific and patent literature, see e.g., Kroll (1993) DNA Cell. Biol., 12:441-53.

Antibody Generation

The invention provides antibodies that specifically bind to the polypeptides of the invention (e.g., the exemplary SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, or SEQ ID NO:8). These antibodies can be used to identify the presence of polypeptides that associated with GCA to aid in its diagnosis and prognosis.

The peptides and polypeptides of the invention can also be used to generate an immune response to generate antibodies for the diagnosis or treatment of GCA (they can also be used

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generate a cellular response for treatment). Thus, they can be administered to both humans and animals. The polypeptides or peptide can be conjugated to another molecule or can be administered with an adjuvant. The coding sequence can be part of an expression cassette or vector capable of expressing the immunogen *in vivo*. (see, *e.g.*, Katsumi (1994) Hum. Gene Ther. 5:1335-9). Methods of producing polyclonal and monoclonal antibodies are known to those of skill in the art and described in the scientific and patent literature, see, *e.g.*, Coligan, Current Protocols in Immunology, Wiley/Greene, NY (1991); Stites (eds.) Basic and Clinical Immunology (7th ed.) Lange Medical Publications, Los Altos, CA ("Stites"); Goding, Monoclonal Antibodies: Principles and Practice (2d ed.) Academic Press, New York, NY (1986); Kohler (1975) Nature 256:495; Harlow (1988) Antibodies, a Laboratory Manual, Cold Spring Harbor Publications, New York.

Antibodies also can be generated *in vitro*, *e.g.*, using recombinant antibody binding site expressing phage display libraries, in addition to the traditional *in vivo* methods using animals. See, *e.g.*, Huse (1989) Science 246:1275; Ward (1989) Nature 341:544; Hoogenboom (1997) Trends Biotechnol. 15:62-70; Katz (1997) Annu. Rev. Biophys. Biomol. Struct. 26:27-45.

Human antibodies can be generated in mice engineered to produce only human antibodies, as described by, e.g., U.S. Patent No. 5,877,397; 5,874,299; 5,789,650; and 5,939,598. B-cells from these mice can be immortalized using standard techniques (e.g., by fusing with an immortalizing cell line such as a myeloma or by manipulating such B-cells by other techniques to perpetuate a cell line) to produce a monoclonal human antibody-producing cell. See, e.g., U.S. Patent No. 5,916,771; 5,985,615.

Nucleic acid sequences (e.g., from cDNA libraries, isolated from human antibody producing mice, etc.) encoding desired antibodies can be cloned and further manipulated. For example, if the antibody is of non-human origin, it can be "humanized" for administration to patients. Methods for making chimeric, e.g., "humanized," antibodies are well known in the art, see e.g., U.S. Patent Nos. 5,811,522; 5,789,554; 5,861,155. Alternatively, recombinant antibodies can also be expressed by transient or stable expression vectors in mammalian, including human, cells and cell lines, as in Norderhaug (1997) J. Immunol. Methods 204:77-87; Boder (1997) Nat. Biotechnol. 15:553-557; see also U.S. Patent No. 5,976,833. CHO cells lines that express "humanized" glycosylation patterns can be particularly useful, see, e.g., U.S. Patent No. 5,272,070.

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In one embodiment, the peptides of the invention are used as a pharmaceutical, immunogenic composition to generate an anti-GCA causative agent response in a human. Alternatively, DNA encoding a polypeptide comprising an immunogenic epitope can be administered as a pharmaceutical. The immunogenic compositions of the invention can generate a humoral (antibody) or cellular immune response.

Specific monoclonal and polyclonal antibodies and antisera will usually bind with a K_D of at least about 1 μ M, preferably at least about 0.1 μ M or better, and most preferably, 0.01 μ M or better.

Diagnosing Giant Cell Arteritis

The invention provides compositions and methods for diagnosing GCA involving the detection of GCA-associated polypeptides and nucleic acids in tissue samples (e.g., vasculitis lesion biopsies) and fluid samples (e.g., serum) from patients. GCA can also be diagnosed by detecting the presence of GCA-associated antibodies in serum or tissue samples from patients. These diagnostic methods are especially useful for the early diagnosis of GCA in the elderly. These procedures can also be used to follow the success of a treatment regimen and make prognoses.

Immunoassays

The invention provides reagents and methods using the polypeptides and peptides of the invention in a variety of antibody based assays. As discussed above, these assays are used to detect human antibodies in serum and tissues from patients to diagnose GCA. As the peptides and polypeptides of the invention can also be used to generate new antibodies for the diagnosis and treatment of GCA, these assays can be used to assess the generation, titer, isotype, etc., the antibodies of the invention.

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Immunological binding methodologies are well known in the art; see also U.S. Patent Nos. 4,366,241; 4,376,110; 4,517,288; 4,837,168; 5,817,470; METHODS IN CELL BIOLOGY Vol. 37,. Antibodies in Cell Biology, Asai, ed. Academic Press, Inc. New York (1993); Sambrook, Stites; Silzel (1998) Clin. Chem. 44:2036-43; Rongen (1997) J. Immunol. Methods 204:105-133.; Hashida (1995) Biotechnol. Annu. Rev. 1:403-51; Bao (1997) J. Chromatogr. B. Biomed. Sci. Appl. 699:463-80; Self (1996) Curr. Opin. Biotechnol. 7:60-5.

In various embodiments of the invention, the polypeptides (or peptides) and antibodies of the invention are immobilized to the "capture" GCA-associated antibodies or polypeptides, respectively. Additional reagent are added to this reaction to detect any specific binding. These so-called "sandwich assays" are commercially useful for detecting or isolating protein or antibodies.

Capture assays utilize a "capture" antibody or protein/ peptide (or nucleic acid in a hybridization variation) that is immobilized to a solid support. After adding a tissue or serum sample, a labeled "signal" molecule is added (typically in solution) that can bind to the captured reagent. The immobilized "capture" antibody or protein and the sample molecule and the "signal" molecule form a "sandwich" complex. To be effective, the signal nucleic acid or protein should not bind substantially with the capture antibody or protein. For example, a peptide or polypeptide of the invention is fixed to a solid support and contacted with a tissue or serum sample. GCA-associated antibodies, if present, will bind to the fixed reagent. Anti-human antibodies are then added to detect "captured" GCA-associated human antibodies. The sensitivity of the assay can be enhanced through use of a signal amplification system that multiplies the signal being detected. For example, an enhancing signal can be generated by attaching fluorescent, radioactive or enzymatic molecules to the anti-human antibody binding reagents (e.g., use of a goat F(ab')2 anti-human IgG-alkaline phosphatase). Use of enzymes and subsequent developing chemicals to enhance signals are commonly called "enzyme-linked immunosorbent assays, or, ELISAs. Other labels can include signal nucleic acid ligands that are bound to labeled antibodies, fluorophores, chemiluminescent agents, additional antibodies (e.g., Abs specific for a complex of a chelating agent and a metallic ion, e.g., a radionuclide) or other ligand-binding molecules (e.g., biotin) that can serve as specific binding pair members for a labeled ligand. Examples of radionuclides include, e.g., as ³H, ¹²⁵I, ³⁵S, ¹⁴C, or ³²P in labeled compositions. Examples of fluorochromes include, e.g., DAPI, fluorescein, Hoechst 33258,

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phycoerythrin (PE), allophycocyanin (APC), R-phycocyanin, B-phycoerythrin, R-phycoerythrin, rhodamine (e.g., tetra-methylrhodamine isothiocyanate—TRITC), Texas red or lissamine. Suitable fluoresceins include, e.g., fluorescein isothiocyanate (FITC), (2-aminoethyl)-thioureido-fluorescein (FTED), fluorescein-thiosemicarbazide (FTSC), (2-aminoethyl)-ureido-fluorescein (FAMCO-E), erythrocin (tetra-iodo-fluorescein), and fluoresceinamine (FAM). A signal from the detectable reagent can be analyzed, for example, using a spectro-photometer to detect color from the chromogenic substrate; a radiation counter to detect radiation (as in an RIA), e.g., a gamma counter for detection of iodine-125; or a fluorometer to detect fluorescence in the presence of light of a certain wavelength. For detection of enzyme-linked reagents, a quantitative analysis can be made using a spectrophotometer, e.g., in the form of a microplate reader (e.g., Molecular Devices, Menlo Park, Calif.) in accordance with the manufacturer's instructions. Assays of the invention can be automated or performed robotically and signals from multiple samples can be detected simultaneously.

Immunoassays can be either competitive or noncompetitive. Noncompetitive immunoassays are assays in which the amount of captured analyte (GCA-associated human antibody) is directly measured. In one "sandwich" assay, for example, the capture agent (a polypeptide or peptide of the invention) can be bound directly to a solid substrate where they are immobilized. These immobilized reagent then capture antibody present in the test sample. The antibody thus immobilized is then bound by a labeling agent, such as a second anti-human antibody reagent bearing a label. Alternatively, the human antibody binding reagent can lack a label, but it may, in turn, be bound by a labeled third reagent (e.g. another antibody), e.g., specific to antibodies of the species from which the second antibody is derived. The second (or third) can be modified with a detectable moiety, such as biotin, to which a another labeled molecule can specifically bind, such as, e.g., enzyme-labeled streptavidin. In a variation of the above, the immobilized reagent can be an antibody of the invention used to capture a GCAassociated polypeptide. The second (soluble) reagent can be, e.g., another GCA-associated polypeptide binding antibody of the invention. Competitive binding assays can also be used. For example, a known amount of labeled human antibody is added to the serum or tissue sample. The sample is then contacted with the capture agent (GCA-associated polypeptides or peptides of the invention). The amount of labeled human antibody bound to the immobilized reagent is

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inversely proportional to the concentration of GCA-polypeptide reactive antibody present in the sample. A hapten inhibition assay is another competitive assay.

These assays can be readily adapted to a variety of different variations, including automated analytical apparatus, of which many are well known in the art, see, e.g., U.S. Patent Nos. 5,981,199; 5,958,202; 5,698,450; 5,648,274; 5,451,504; 5,424,220; 5,395,754; 5,175,086. See also U.S. Patent Nos. 5,932,429; 5,780,319, 5,629,167; describing detecting and quantitation of specific human antibodies from tissue or serum samples.

The present invention also provides methods for Western blot (immunoblot) analysis to detect and/or quantify the presence of a protein in a sample. The technique generally comprises separating sample proteins by gel electrophoresis on the basis of molecular weight, transferring the separated proteins to a suitable solid support, (such as a nitrocellulose filter, a nylon filter, or derivatized nylon filter), and incubating the sample with the antibodies that specifically bind, e.g., GCA-associated polypeptides. These antibodies may be directly labeled or alternatively may be subsequently detected using labeled antibodies (e.g., labeled sheep anti-mouse antibodies.

The compositions and methods of the invention are also compatible with other assay formats, including liposome immunoassays (LIA) (Rongen (1997) *J. Immunol. Methods* 204:105-133), in which liposomes designed to bind specific molecules (*e.g.*, antibodies or polypeptides) and release encapsulated reagents or markers are employed. The released chemicals can be detected using standard techniques (*see*, *e.g.* Monroe (1986) *Amer. Clin. Prod. Rev.* 5:34).

Detecting GCA-Associated Nucleic Acid

GCA can also be diagnosed by detecting the presence of GCA-associated nucleic acid (e.g., the exemplary sequences of the invention, SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:5; SEQ ID NO:7; and SEQ ID NO:9 through 14.

One method for evaluating the presence or absence of GCA-associated DNA in a sample involves a Southern transfer. Briefly, the nucleic acid sample, such as digested DNA (e.g., genomic, cDNA) or mRNA, is run on agarose slab or polyacrylamide gels in buffer and transferred to membranes. Hybridization is carried out using nucleic acid probes. Nucleic acid

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probes can be, e.g., 10 to 20 to 30 or more bases or longer in length (see Sambrook for methods of selecting nucleic acid probe sequences for use in nucleic acid hybridization). Both quantitative and qualitative determination of the presence or absence of DNA or RNA encoding protein can be performed. Similarly, a Northern transfer can be used for the detection of GCA-associated mRNA. For example, mRNA is isolated from a given cell sample using an acid guanidinium-phenol-chloroform extraction method. The mRNA is then electrophoresed to separate the mRNA species and the mRNA is transferred from the gel to a nitrocellulose membrane. As with the Southern transfers, probes, such as labeled probes or PCR amplification products can be used to identify the presence or absence of GCA-associated nucleic acid.

Typically, oligonucleotide probes are labeled signal nucleic acids that are used to detect hybridization. Complementary probe nucleic acids or signal nucleic acids can be labeled by any means used to detect the presence of hybridized polynucleotides (typically, the same as labels on antibodies, as discussed above). Methods of detection can use labels for autoradiography or autofluorography, such as, e.g., ³H, ¹²⁵I, ³⁵S, ¹⁴C, or ³²P-labeled probes or the like. Other labels include signal nucleic acid ligands that are bound to labeled antibodies, fluorophores, chemiluminescent agents, enzymes, and antibodies that can serve as specific binding pair members for a labeled ligand, as described above for antibody sandwich (e.g., ELISA) assays. Amplification (e.g., PCR) techniques (described above) can also be used to directly detect GCA-associated nucleic acids *in situ* or to amplify signals in capture assays.

Nucleic acid hybridization assays can also be performed in an array-based format; and the invention provides arrays comprising the nucleic acids of the invention. Arrays are a multiplicity of different "probe" or "target" nucleic acids (or other compounds) are hybridized against a target nucleic acid. In this manner a large number of different hybridization reactions can be run essentially "in parallel". This provides rapid, essentially simultaneous, evaluation of a wide number of reactants. Variations on arrays include, e.g., spectral imaging methods aimed at detecting and analyzing fluorescent *in situ* hybridizations employing numerous chromosome paints and/or loci specific probes, each labeled with a different fluorophore or a combination of fluorophores, see, e.g., U.S. Patent No. 5,936,731. Methods of performing hybridization reactions in array based formats are well known to those of skill in the art, e.g., Jackson (1996) *Nature Biotechnology* 14:1685; Chee, *Science* 274:610 (1995); U.S. Patent Nos: 6,004,755; 6,004,752; 5,631,134.

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An alternative means for determining the level of expression of a gene encoding a protein is *in situ* hybridization. *In situ* hybridization assays are well known and are generally described in Angerer (1987) *Methods Enzymol* 152:649. In an *in situ* hybridization assay, cells are fixed to a solid support, typically a glass slide or analyzed by a fluorescence activated cell sorter (FACS). If DNA is to be probed, the cells are typically denatured with heat or alkali. The cells are then contacted with a hybridization solution at a moderate temperature to permit annealing of labeled probes specific to the nucleic acid sequence encoding the protein. The probes are typically labeled, *i.e.*, with radioisotopes or fluorescent reporters. See also U.S. Patent No. 5,583,016; FISH fluorescence *in situ* hybridization, as described by Macechko (1997) *J Histochem Cytochem* 45:359-363; Raap (1995) *Hum Mol Genet* 4(4), 529-534.

Treating Giant Cell Arteritis

The invention provides compositions and methods for the prevention, treatment or amelioration of GCA. The therapeutic compositions of the invention can be administered as nucleic acids that can hybridize to GCA-causative nucleic acids *in vivo* (e.g., a nucleic acid from a GCA-causative microorganism) or an antibody of the invention reactive with a GCA-causative polypeptide.

Inhibitory Oligonucleotides

One particularly useful set of inhibitors provided by the present invention includes oligonucleotides which are able to either bind mRNA encoding GCA-causative nucleic acids or to their corresponding genes, in either case preventing or inhibiting the production of functional proteins involved in the pathogenesis of GCA. The inhibitory association can be though sequence specific hybridization to another nucleic acid or by general binding, as in an aptamer. In another embodiment, RNA can be targeted for cleavage by RNAase P from eukaryotic cells (e.g., human RNAase P) using a suitably designed antisense oligo-ribonucleotides as "external guide sequences" to form a hybrid with the target RNA, thereby creating a substrate for cleavage by RNAase P *in vitro*; see e.g., U.S. Patent No. 5,624,824. One useful class of inhibitors includes oligonucleotides which cause inactivation or cleavage of mRNA. For example, the oligonucleotide is chemically modified or has enzyme activity which causes such cleavage of the

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mRNA, such as a ribozyme. Screening for effective oligonucleotides with the desired activity can be done by routine methods. Another useful class of inhibitors includes oligonucleotides which bind polypeptides. Double- or single-stranded DNA or single-stranded RNA molecules that bind to specific polypeptides targets are called "aptamers." The specific oligonucleotide-polypeptide association may be mediated by electrostatic interactions. For example, aptamers specifically bind to anion-binding exosites on thrombin, which physiologically binds to the polyanionic heparin (Bock (1992) *Nature* 355:564-566). Screening for GCA polypeptide-binding aptamers also can be done by routine methods.

GCA-causative activity also can be inhibited by targeting the mRNA *in vivo* with antisense oligonucleotides. In some situations, naturally occurring nucleic acids used as antisense oligonucleotides may need to be relatively long (18 to 40 nucleotides) and present at high concentrations. A wide variety of synthetic, non-naturally occurring nucleotide and nucleic acid analogues are known which can address this potential problem. For example, protein-nucleic acids (PNAs) containing non-ionic backbones, such as N-(2-aminoethyl) glycine units can be used. Antisense oligonucleotides having phosphorothioate linkages can also be used, as described in WO 97/03211; WO 96/39154; Mata (1997) *Toxicol Appl Pharmacol* 144:189-197; Antisense Therapeutics, ed. Agrawal (Humana Press, Totowa, NJ, 1996). Antisense oligonucleotides having synthetic DNA backbone analogues provided by the invention can also include phosphoro-dithioate, methylphosphonate, phosphoramidate, alkyl phosphotriester, sulfamate, 3'-thioacetal, methylene(methylimino), 3'-N-carbamate, and morpholino carbamate nucleic acids. As with sequence effective for ribozyme activity, screening for effective antisense sequences can be done by routine screening, see, e.g., U.S. Patent No. 5,580,967; 6,013,447.

Combinatorial chemistry methodology also can be used to create vast numbers of oligonucleotides that can be rapidly screened for specific oligonucleotides that have appropriate binding affinities and specificities toward any target, such as the GCA-causative proteins of the invention; see, e.g., U.S. Patent Nos. 5,880,972; 5,792,431; 5,529,756; 5,503,805.

Inhibitory Ribozymes

Ribozymes are enzymatic RNA molecules capable of catalyzing the specific cleavage of RNA. The mechanism of ribozyme action involves sequence-specific hybridization of the ribozyme molecule to complementary target RNA, followed by endonucleolytic cleavage.

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Within the scope of the invention are engineered hammerhead motif ribozyme molecules that can specifically and efficiently catalyze endonucleolytic cleavage of the sequences encoding GCA-causative polypeptides.

Specific ribozyme cleavage sites within any potential RNA target are initially identified by scanning the target molecule for ribozyme cleavage sites which include the following sequences, GUA, GUU and GUC. Once identified, short RNA sequences of between 15 and 20 ribonucleotides corresponding to the region of the target gene containing the cleavage site may be evaluated for secondary structural features which may render the oligonucleotide inoperable. The suitability of candidate targets may also be evaluated by testing accessibility to hybridization with complementary oligonucleotides using ribonuclease protection assays.

Ribozymes act by binding to a target RNA through the target RNA binding portion of a ribozyme which is held in close proximity to an enzymatic portion of the RNA that cleaves the target RNA. Thus, the ribozyme recognizes and binds a target RNA through complementary base-pairing, and once bound to the correct site, acts enzymatically to cleave and inactivate the target RNA. Cleavage of a target RNA in such a manner will destroy its ability to direct synthesis of an encoded protein if the cleavage occurs in the coding sequence. After a ribozyme has bound and cleaved its RNA target, it is typically released from that RNA and so can bind and cleave new targets repeatedly.

The enzymatic ribozyme RNA molecule is able to cleave RNA and thereby inactivate a target RNA molecule. The complementarity functions to allow sufficient hybridization of the enzymatic ribozyme RNA molecule to the target RNA for cleavage to occur. Complementarity as low as 50-75% may also be employed. The present invention provides ribozymes targeting any portion of the coding region for a GCA-causative gene that cleave mRNA in a manner that will inhibit the translation of the mRNA.

The enzymatic ribozyme RNA molecule can be formed, e.g., in a hammerhead motif, but may also be formed in the motif of a hairpin, hepatitis delta virus, group I intron or RNaseP-like RNA (in association with an RNA guide sequence). Examples of such hammerhead motifs are described by Rossi (1992) *Aids Research and Human Retroviruses* 8:183; hairpin motifs by Hampel (1989) *Biochemistry* 28:4929, and Hampel (1990) *Nuc. Acids Res.* 18:299; the hepatitis delta virus motif by Perrotta (1992) *Biochemistry* 31:16; the RNaseP motif by Guerrier-Takada (1983) *Cell* 35:849; and the group I intron by Cech U.S. Pat. No. 4,987,071.

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Preparation and use of inhibitory nucleic acids

Antisense molecules and ribozymes of the invention may be prepared by any method known in the art for the synthesis of RNA molecules. These include techniques for chemically synthesizing oligonucleotides such as solid phase phosphoramidite chemical synthesis.

Alternatively, RNA molecules may be generated by *in vitro* and *in vivo* transcription of DNA sequences encoding the GCA-associated polypeptides of the invention. Such DNA sequences may be incorporated into a wide variety of vectors with suitable RNA polymerase promoters such as T7 or SP6. Alternatively, antisense cDNA constructs that synthesize antisense RNA constitutively or inducibly can be introduced into cell lines, cells or tissues.

RNA molecules can be modified to increase intracellular stability and half-life. Possible modifications include, but are not limited to, the addition of flanking sequences at the 5' and/or 3' ends of the molecule or the use of phosphorothioate or 2' O-methyl rather than phosphodiesterase linkages within the backbone of the molecule. This concept is inherent in the production of PNAs and can be extended in all of these molecules by the inclusion of nontraditional bases such as inosine, queosine and wybutosine as well as acetyl-, methyl-, thio- and similarly modified forms of adenine, cytidine, guanine, thymine, and uridine which are not as easily recognized by endogenous endonucleases.

Triplex DNA can also be used to inhibit DNA transcription and replication, generate site-specific mutations, cleave DNA, and induce homologous recombination. See, e.g., Havre (1993) *J. Virology* 67:7324-7331; Scanlon (1995) *FASEB J.* 9:1288-1296; Giovannangeli (1996) *Biochemistry* 35:10539-10548; Chan (1997) *J. Mol. Medicine (Berlin)* 75: 267-282. Triple helix DNAs can be used to target the same sequences identified for antisense regulation.

Methods for introducing vectors into cells or tissues include those methods that are suitable for *in vivo*, *in vitro* and *ex vivo* therapy, as described below.

Antibodies for treating GCA

In addition to providing antibodies reactive with GCA-associated polypeptides useful for diagnosing GCA, the invention also provides antibodies reactive with GCA-causative polypeptides for the amelioration, prevention or treatment of GCA. Administration of antibodies

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can be as polypeptides or as nucleic acids encoding antibodies; such methods and protocols are well known in the art.

Formulation and Administration Pharmaceuticals

The invention provides antibodies (directed to GCA-causative proteins, e.g., SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:8), vectors or oligonucleotides (e.g., inhibitory nucleic acids, e.g., antisense) with pharmaceutically acceptable carrier (excipient) to form a pharmacological composition. The nucleic acids and vectors can be formulated as pharmaceuticals for the transfer of nucleic acids into cells *in vitro* or *in vivo*. The pharmaceutical composition of the invention can further comprise other active agents, including other recombinant viruses, plasmids, naked DNA or pharmaceuticals (e.g., anti-inflammatory agents).

These pharmaceuticals can be administered by any means in any appropriate formulation. Routine means to determine drug regimens and formulations to practice the methods of the invention are well described in the patent and scientific literature, and some illustrative examples are set forth below. For example, details on techniques for formulation, dosages, administration and the like are well described in the scientific and patent literature, see, *e.g.*, the latest edition of Remington's Pharmaceutical Sciences, Maack Publishing Co, Easton PA.

Pharmaceutically acceptable carriers can contain a physiologically acceptable compound that acts, e.g., to stabilize the composition or to increase or decrease the absorption of the agent and/or pharmaceutical composition. Physiologically acceptable compounds can include, for example, carbohydrates, such as glucose, sucrose, or dextrans, antioxidants, such as ascorbic acid or glutathione, chelating agents, low molecular weight proteins, compositions that reduce the clearance or hydrolysis of any co-administered agents, or excipients or other stabilizers and/or buffers. Detergents can also used to stabilize the composition or to increase or decrease the absorption of the pharmaceutical composition (see infra for exemplary detergents).

Other physiologically acceptable compounds include wetting agents, emulsifying agents, dispersing agents or preservatives that are particularly useful for preventing the growth or action of microorganisms. Various preservatives are well known, e.g., ascorbic acid. One skilled in the art would appreciate that the choice of a pharmaceutically acceptable carrier, including a physiologically acceptable compound depends, e.g., on the route of administration of the

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adenoviral preparation and on the particular physio-chemical characteristics of any coadministered agent.

The compositions for administration will commonly comprise a buffered solution comprising nucleic acid (e.g., vector) or antibody in a pharmaceutically acceptable carrier, e.g., an aqueous carrier. A variety of carriers can be used, e.g., buffered saline and the like. These solutions are sterile and generally free of undesirable matter. These compositions may be sterilized by conventional, well-known sterilization techniques. The compositions may contain pharmaceutically acceptable auxiliary substances as required to approximate physiological conditions such as pH adjusting and buffering agents, toxicity adjusting agents and the like, for example, sodium acetate, sodium chloride, potassium chloride, calcium chloride, sodium lactate and the like. The concentration of active agent in these formulations can vary widely, and will be selected primarily based on fluid volumes, viscosities, body weight and the like in accordance with the particular mode of administration selected and the patient's needs.

Determining Dosing Regimens

The pharmaceutical formulations of the invention can be administered in a variety of unit dosage forms, depending upon the particular condition or disease, the general medical condition of each patient, the method of administration, and the like. In one embodiment, the concentration of vectors (e.g., therapeutic virus or plasmids) in the pharmaceutically acceptable excipient is between about 10³ to about 10¹⁸ or between about 10⁵ to about 10¹⁵ or between about 10⁶ to about 10¹³ particles/vectors per mL in an aqueous solution. Details on dosages are well described in the scientific and patent literature, see, *e.g.*, the latest edition of Remington's Pharmaceutical Sciences; Sterman (1998) Hum. Gene Ther. 9:1083-1092; Smith (1997) Hum. Gene Ther. 8:943-954.

The exact amount and concentration of antibody or vector or oligonucleotide and the amount of formulation in a given dose, or the "therapeutically effective dose" is determined by the clinician, as discussed above. The dosage schedule, *i.e.*, the "dosing regimen," will depend upon a variety of factors, e.g., the stage and severity of the GCA, and the general state of the patient's health, physical status, age and the like. The state of the art allows the clinician to determine the dosage regimen for each individual patient and, if appropriate, concurrent disease

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or condition treated. For example, adenovirus has been safely used for many years for human vaccines; see, e.g., Horwitz (1990) supra; Straus (1984) supra; Haj-Ahmad (1986) J. Virol., 57:267); Ballay (1985) EMBO, 4, 3861 (1985); PCT patent application WO 94/17832). Human adenoviruses have been used in humans as in vivo gene delivery vehicles (Graham & Prevec in New Approaches to Immunological Problems, Ellis (ed), Butterworth-Heinemann, Boston, Mass., pp. 363-390 (1992); Ragot (1993) Nature 361:647-650 (1993); Kozarsky (1993) Curr. Opin. Genet. Dev. 3:499-503); U.S. Patent No. 5,981,225. These illustrative examples can also be used as guidance to determine the dosage regiment, i.e., dose schedule and dosage levels administered when practicing the methods of the invention.

Single or multiple administrations of antibody, vector or oligonucleotide formulation can be administered, depending on the dosage and frequency as required and tolerated by the patient. Thus, one typical dosage for regional (e.g., IP or intrathecal) administration is between about 0.5 to about 50 mL of a formulation with about 10^{13} vectors/ particles per mL. In an alternative embodiment, dosages are from about 5 mL to about 20 mL are used of a formulation with about 10^9 vectors/particles per mL. Lower dosages can be used, such as is between about 1 mL to about 5 mL of a formulation with about 10^6 vectors/particles particles per mL. Based on objective and subjective criteria, as discussed herein, any dosage can be used as required and tolerated by the patient.

The exact concentration of antibodies, vector or oligonucleotide, the amount of formulation, and the frequency of administration can also be adjusted depending on the levels of *in vivo* (e.g., *in situ*) gene expression and vector retention after an initial administration.

Routes of Delivery

The pharmaceutical compositions of the invention (e.g., therapeutic antibodies, vectors or antisense oligonucleotides) can be delivered by any means known in the art systemically (e.g., intravenously), regionally, or locally (e.g., intravenously), parenteral, intravenously), parenteral, intravenously, parenteral, intravenously, parenteral, intravenously, topical, oral, or local administration, as subcutaneous, intravenously, by aerosol) or transmucosal (e.g., buccal, bladder, vaginal, uterine, rectal, nasal mucosa), intratumoral (e.g., transdermal application or local injection). For example, intraversal injections

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can be used to have a 'regional effect," e.g., to focus on a specific organ (e.g., brain, liver, spleen, lungs). For example, intra-hepatic artery injection or intra-carotid artery injection. If it is desired to deliver the preparation to the brain, it can be injected into a carotid artery or an artery of the carotid system of arteries (e.g., occipital artery, auricular artery, temporal artery, cerebral artery, maxillary artery, etc.).

Therapeutic antibodies, vectors or other nucleic acids of the present invention, alone or in combination with other suitable components can be made into aerosol formulations to be administered via inhalation. These aerosol formulations can be placed into pressurized acceptable propellants, such as dichlorodifluoromethane, propane, nitrogen, and the like. They also may be formulated as pharmaceuticals for non-pressured preparations such as in a nebulizer or an atomizer. Typically such administration is in an aqueous pharmacologically acceptable buffer as described above. Delivery to the lung can be also accomplished, e.g., by use of a bronchoscope.

Additionally, the therapeutic compositions employed in the present invention may be made into suppositories by mixing with a variety of bases such as emulsifying bases or water-soluble bases. Formulations suitable for vaginal administration may be presented as pessaries, tampons, creams, gels, pastes, foams, or spray formulas.

The pharmaceutical formulations of the invention can be presented in unit-dose or multi-dose sealed containers, such as ampules and vials, and can be stored in a freeze-dried (lyophilized) condition requiring only the addition of the sterile liquid excipient, for example, water, for injections, immediately prior to use. Extemporaneous injection solutions and suspensions can be prepared from sterile powders, granules, and tablets.

Therapeutic compositions can also be administered in a lipid formulation, more particularly either complexed with liposomes to for lipid/nucleic acid complexes (e.g., as described by Debs and Zhu (1993) WO 93/24640; Mannino (1988) supra; Rose, U.S. Pat No. 5,279,833; Brigham (1991) WO 91/06309; and Felgner (1987) supra) or encapsulated in liposomes, as in immunoliposomes directed to specific cells. It will be appreciated that such lipid formulations can also be administered topically, systemically, or delivered via aerosol.

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Kits

The invention provides kits that contain the vectors or pharmaceutical compositions of the invention. The kits can contain, e.g., vectors able to produce antisense sequences to inhibit causative agents of GCA. The kit can contain instructional material teaching methodologies, e.g., means to diagnose GCA or biopsy arteritis lesions. Kits containing pharmaceutical preparations (e.g., vectors, nucleic acids) can include directions as to indications, dosages, routes and methods of administration, and the like.

It is understood that the examples and embodiments described herein are for illustrative purposes only and that various modifications or changes in light thereof will be suggested to persons skilled in the art and are to be included within the spirit and purview of this application and scope of the appended claims.

EXAMPLES

The following examples are offered to illustrate, but not to limit the claimed invention.

Example 1: Isolation and Characterization of Sequences Associated With GCA

The following example describes the isolation and identification of the novel sequences of the invention by subtractive hybridization of nucleic acid of normal tissue from nucleic acid of GCA lesions.

Because of advances in techniques of representational difference analysis (RDA), this method was chosen to detect unique sequences in samples of normal versus GCA-associated DNA. This method can identify sequences from pathogenic organisms, latent or active, in tissue lesion samples in addition to genomic losses or rearrangements. After isolation, nucleic acid from an involved site is compared to DNA obtained from uninvolved tissue. RDA represents a powerful tool in which differences between complex genomes can be identified. Recent modifications in the technique permit the analysis of minute samples from small cell numbers.

A genomic (rather than RNA expression) strategy was selected in order to decrease the accrual of inflammation-associated host genes, such as cytokines or T cell associated polypeptides, as T cell receptors (TCRs). Genomic RDA systematically excludes the majority of

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"host" (i.e., patients') genes and permits identification of sequences from any GCA-associated microorganism with a DNA phase to its life-cycle.

Like other subtractive strategies, RDA critically depends on optimal positive ("GCA-involved" or "lesional" or "tester") and negative ("normal" or "non-lesional" or "driver") starting material. Since temporal artery lesions in GCA are discontinuous, the ideal pair of tester and driver areas can be present in biopsies from single individuals. A microscopic-scale laser microdissection isolation method was used to obtain homogeneous lesional and non-lesional samples. This is a flexible and direct method for isolating microscopic-scale samples for molecular analysis. Laser-capture microdissection was used to isolate DNA from archival pathology specimens of GCA-positive arteries from both histopathologically involved and uninvolved areas. Genomic RDA was performed on archival, GCA-positive temporal arteries that were snap-frozen. These specimens provided a source of high quality genomic DNA. Specimens that were selected for RDA unequivocally demonstrated pathologic evidence of GCA.

Thirty sequentially cut tissue sections (10 µm each) were placed on non-charged microscope slides, fixed, dehydrated, and stained with hematoxylin and eosin. GCA inflammatory lesions were identified and dissected using a Laser Capture Microdissecting MicroscopeTM (Arcturus Engineering, Inc., Mountain View, CA). Approximately 500 cells including giant cells and inflammatory infiltrate were dissected from each GCA specimen. DNA isolated from these cells represented the "tester" population. Approximately 500 cells were microdissected from a subsequent section with no histopathologic evidence of GCA lesions. The DNA isolated from these cells represent the "driver" sample in the RDA.

DNA was extracted from tester and driver samples and prepared for RDA according to Michiels (1998) supra. Briefly, isolated cells were incubated overnight at 37°C in proteinase K and restriction digested with BamH1. The driver and tester DNA were ligated to amplimers RBAM14 and RBAM24 and PCR amplified as described by Lisitsyn (1993) supra and Michiels (1998) supra. RDA was performed using the lesional DNA as the tester and non-lesional DNA as the driver in consecutive rounds of hybridization/ amplification to obtain sequences represented solely in the lesional DNA population.

The fourth round of RDA yielded two prominent bands. This entire region of the gel encompassing these prominent bands was excised and the DNA was extracted and cloned into a

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TA vector (Invitrogen, San Diego, CA). Bacteria were transformed with the vector and 100 individual clones containing insert were isolated. The 100 clones were examined for insert by PCR amplification. Eighty percent of the clones contained insert. Analysis of these clones revealed multiple inserts of unique size.

Priority candidate genes within the two RDA libraries were identified using several steps. First, duplicate clones were excluded by DNA fragment size and by 4-cutter restriction pattern. Second, unique cloned inserts were analyzed by automated sequencing. Third, the sequences were used to search the National Institutes of Health non-redundant database with the NCBI BLAST program (BLOSUM 62 matrix, TBLASTX (version 1.4.11, Nov-97) as described by Alschul (1996) Methods Enzymol. 266:460-480. Alignments were performed using the CLUSTAL W Multiple Sequence Alignment Program (version 1.7, June, 1997).

One criterion to for selecting which genes would first be translated into recombinant and used to analyze for the presence of human antibodies in serum from GCA patients was relative homology to sequences of known microbial origin. A total of eleven unique DNA sequences were obtained in the first two RDAs using two different GCA+ arterial specimens. Sequence identity analysis showed that four of the unique sequences, GCA 1, GCA 5, GCA 14, and GCA 17, may be distantly related to known microbial sequences. Thus, these were the first sequences selected for further characterization.

Example 2: Serum Antibodies From GCA Patients Specifically Bind to Polypeptides of the **Invention**

The following example demonstrates that antibodies in the serum of GCA patients specifically bind to exemplary polypeptides of the invention.

Validation of GCA-associated gene segments relies, in part, on functional evaluations of either the presence of disease-specific immune responses against the associated candidate protein sequence or disease-specific expression of the gene sequence or product. In order to proceed with analysis of the novel sequences of the invention, open reading frames from these sequences were analyzed and expression constructs produced.

These unique sequences GCA 1, GCA 5, GCA 14, and GCA 17, as identified in Example 1, were analyzed for open reading frames (ORF). A total of five ORFs from four sequences were identified. These were used to construct fusion proteins to identify the presence

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of anti- GCA 1, GCA 5, GCA 14, and GCA 17 antibodies in human GCA patients. The sequence of the fusion proteins is set forth, above.

GST fusion proteins were produced using these five identified ORFs from the clones identified by genomic RDA. Each ORF was amplified using sequence specific primers designed to produce an in-frame product when cloned into the multiple cloning site of an expression vector, pGEX-KG, as described by Guan (1991) Anal. Biochem. 192:262-267. The BL21 strain of E. coli (Stratagene, San Diego, CA) was transformed with the desired pGEX-GCA construct. Individual colonies were selected, grown, and its sequence confirmed as the correct insertion of one in-frame fragment copy. Individual colonies were selected, grown, and expression of fusion protein was induced with IPTG using standard techniques. GST (glutathione S-transferase)-GCA fusion proteins were produced and purified using the GST Gene fusion System (Pharmacia Biotech) and manufacturer's instructions. Briefly, the bacterial cells were lysed by sonication and the soluble GST fusion proteins allowed to bind to a glutathione Sepharose 4B column. Purified GST-fusion proteins were eluted from the column and protein concentration determined by the Bradford method. Several of the GST-GCA fusion proteins were insoluble and localized to the inclusion bodies. These proteins were retrieved by isolation of the inclusion body followed by Solubilization of the recombinant proteins in the inclusion bodies. Purity was estimated at greater than 90% by SDS-PAGE and protein (Coomassie) stain. Reactivity of the expected product was determined by Western blot analysis using a commercially available anti-GST antibody (Molecular Probes). The GST-GCA fusion proteins were then evaluated for GST reactivity to human serum.

The protocol for large-scale GST-fusion protein purification was: Day 1:

1. Pick colony from plate (LB-agar plate with 0.05mg amp/ml) plated from glycerol stocks. Grow in 5ml LB-amp o/n shaking at 37°C.

<u>Day 2</u>:

2. Preparation of Beads (in morning): 950mg of G4510 Sigma Glutathione Agarose Beads in 20ml PBS. Rock for 2 hrs at room temperature (r.t.). Store at 4°C. Warm to room temperature to prepare the column.

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- 3. Preparation of (1ml) Columns): Prepare quiagen column with slurry to produce a 1 ml packed column. Equilibrate column in Bugbuster + 125µl 1M DTT + 2 complete tablets (Boehringer) + 50µl 250mM PMSF(50mM final concentration).
- 4. In morning transfer 5ml culture into 600ml LB-amp and continue shaking at 37°C. Grow until OD reading (at 600 nm) is 0.6-0.8.
- 5. Induce w/ 1 μ l of 100mM IPTG per ml of culture. at r.t. shaking vigorously for ~1 ½ hrs.
- 6. Centrifuge at 5,000xg for 10 min.Decant and remove as much liquid as possible, leaving pellet.
- 7. Resuspend pellets in Bugbuster (Novagen, 70584-4) (1/25 th volume of the original culture volume). Prepare 25ml Bugbuster + 125μl of 1M DTT (5mM DTT final), 2 complete tabs (Boehringer) +50μ l of 250mM PMSF (0.5mM final PMSF conc.); store on ice. Add 12 ml of Bugbuster mix to each pellet and vortex.
- 8. Incubate centrifuge tubes at r.t. shaking at slow setting for 10 min.
- 9. Transfer each 12ml soup to a 30ml small autoclaved centrifuge tube. Spin at 16,000xg for 20 min. at 4°C.
- 10. Apply supernatant to prepared columns. Wash Buffers: Wash 1: 50mM Tris-Cl, pH 7.5; 300mM NaCl; 10mM EDTA; complete tablet 0.5 mM PMSF. Wash 2: Above buffer, without NaCL. Elution Buffer: 50mM Tris-Cl, pH 8, 25mM reduced glutathione.
- 11. Uncap columns and collect flow thru.
- 12. Wash each column in 2x6ml wash 1
- 13. Wash each column in 2x6ml wash 2
- 14. Cap columns and add 2ml of elution buffer. (Can store in 4°C at this stage.)
- 15. Elute in 500µl, 1ml, and finally 2ml of elution buffer. Check OD of each.
- 16. Quantify protein concentration by Bradford method
- 25 16. Store eluted protein fractions in -20°C or -80°C.

GST Fusion Protein ELISA Testing

ELISA analyses were performed for primary detection of human serum antibodies with specificity for the GST-GCA fusion proteins. The GST fusion proteins (purified as noted above) and a GST protein were purified as a control. Microtiter plates were coated with a GST-GCA

fusion protein or GST control; 1 μ g/well in 50 μ l borate coating buffer (5.15 g of boric acid, 3.65 g NaCl, per 500 ml, pH 8.5; washed, 3X with 0.05% Tween20 –PBS, and blocked with 0.05% Tween20 –PBS.

Human sera was tested in triplicate at multiple dilutions in 0.05% Tween20 –PBS. Reactivity was detected with an alkaline-phosphatase-labeled goat anti-human IgG and developed with Sigma 104 phosphatase substrate. Absorbances was measured at 405 nm with a Biorad ELISA reader and Macintosh analytic software. OD values of nonspecific binding of sera to GST alone was subtracted from the raw values of binding to the GST-GCA fusion proteins in order to determine specific absorbances.

Figure 1 illustrates data from a representative ELISA using the fusion protein GCA1b-GST to test human serum for the presence of anti-GCA1b reactive antibodies. Various dilutions of human serum from a GCA+ patient and GCA- individual were used, as shown in Figure 1. Specific binding of human antibodies to GCA1b was detected in the patient's serum (note enhanced serum binding to the GCA+ serum, as compared to GCA-, or "normal" serum).

Figure 2 illustrates data from a representative ELISA using the fusion protein GCA17-GST to test human serum for the presence of anti- GCA17 reactive antibodies. Various dilutions of human serum from a GCA+ patient and a GCA- individual were used, as shown in Figure 2. Specific binding of human antibodies to GCA17 was detected in the patient's serum (note enhanced serum binding to the GCA+ serum, as compared to GCA-, or "normal" serum).

Figure 3 illustrates a representative experiment using four exemplary GCA-GST fusion proteins, GCA 1a-GST, GCA 1b-GST, GCA14-GST, and GCA17-GST. These fusion proteins were used to detect serum IgG in sera from 10 GCA+ patients and 10 GCA- individuals. All sera were individually tested at multiple dilutions. The graph demonstrates the mean OD observed after 30 minutes of detection and the standard deviation for binding of each of the fusion proteins to the sera at 1/100 dilution.

These data demonstrate that the peptides and polypeptides of the invention can be used to identity antibodies present in the serum of GCA+ patients. Accordingly, these compositions and associated methods of the invention can be used to diagnose, and, importantly, to screen for, the presence or predisposition to GCA.

These data also demonstrate that the peptides and polypeptides of the invention can be used to generate additional antibodies (the antibodies of the invention) to further identify the

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predisposition to or presence of GCA. These antibodies can also be used to treat GCA, as described above.

Because these data demonstrate that that the peptides and polypeptides of the invention are associated with GCA, they also demonstrate that the nucleic acids of the invention also can be used to diagnose GCA. As these nucleic acids also encode polypeptides possibly essential to the GCA-causative agent, these data also demonstrate that the nucleic acids of the invention also can be used to treat GCA, as described above.